

Review

Innovative Pulses for Western European Temperate Regions: A Review

Alicia Ayerdi Gotor ^{1,*}  and Elisa Marraccini ^{2,3} 

¹ Agro-Ecologie, Hydrogéochemie, Milieux & Ressources UP 2018.C101, SFR Condorcet FR CNRS 3417, UniLaSalle, 60026 Beauvais, France

² Innovation, Territoire, Agriculture & Agro-Industrie, Connaissance et Technologie UP 2018.C102, SFR Condorcet FR CNRS 3417, UniLaSalle, 60026 Beauvais, France; elisa.marraccini@uniud.it

³ Department of Agricultural, Food, Environmental and Animal Sciences, University of Udine, 33100 Udine, Italy

* Correspondence: Alicia.ayerdi-gotor@unilasalle.fr; Tel.: +33-(3)-4406-2549

Abstract: In Europe, there is an increasing interest in pulses both for their beneficial effects in cropping systems and for human health. However, despite these advantages, the acreage dedicated to pulses has been declining and their diversity has reduced, particularly in European temperate regions, due to several social and economic factors. This decline has stimulated a political debate in the EU on the development of plant proteins. By contrast, in Southern countries, a large panel of minor pulses is still cropped in regional patterns of production and consumption. The aim of this paper is to investigate the potential for cultivation of minor pulses in European temperate regions as a complement to common pulses. Our assumption is that some of these crops could adapt to different pedoclimatic conditions, given their physiological adaptation capacity, and that these pulses might be of interest for the development of innovative local food chains in an EU policy context targeting protein autonomy. The research is based on a systematic review of 269 papers retrieved in the Scopus database (1974–2019), which allowed us to identify 41 pulses as candidate species with protein content higher than 20% that are already consumed as food. For each species, the main agronomic (e.g., temperature or water requirements) and nutritional characteristics (e.g., proteins or antinutritional contents) were identified in their growing regions. Following their agronomic characteristics, the candidate crops were confronted with variability in the annual growing conditions for spring crops in Western European temperate areas to determine the earliest potential sowing and latest harvest dates. Subsequently, the potential sum of temperatures was calculated with the Agri4cast database to establish the potential climatic suitability. For the first time, 21 minor pulses were selected to be grown in these temperate areas and appear worthy of investigation in terms of yield potential, nutritional characteristics or best management practices.

Keywords: grain legumes; crop diversification; sustainable intensification; growing degree days



Citation: Ayerdi Gotor, A.; Marraccini, E. Innovative Pulses for Western European Temperate Regions: A Review. *Agronomy* **2022**, *12*, 170. <https://doi.org/10.3390/agronomy12010170>

Academic Editor: Marten Sørensen

Received: 14 December 2021

Accepted: 5 January 2022

Published: 11 January 2022

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

There is an increasing debate about the importance of plant-based proteins and the diversification of protein sources for food or feed [1]. Plant-based proteins originate in several botanical families but are mainly concentrated in legumes. Despite the importance of grain legume, their production and consumption are declining worldwide [2,3], whereas the acreage dedicated to soybean is continuously increasing due to the intensification of livestock production [3]. Although grain legume diversity is very high from a worldwide perspective [4], these species often have local patterns of production and consumption in several developed and developing countries [5]. Worldwide grain legumes production increased by 15% between 2016 and 2017. This increase in production occurred in each continent, except America where production remained stable. Soybean production also increased during this period. This increase was mainly due to the largest producers, North

and South America. Based on the most recent available FAOSTAT data [6], there are four main pulses cultivated worldwide. Beans (all *Phaseolus* species included) were the most produced in 2019 (Table 1), half of which were produced in Asia. Peas are produced in Europe and in North America; however, 83% of chickpeas production (including the species *Cicer arietinum*) was harvested in Asia. Beyond these four common grain legumes, cowpea production reached 8.9 million tons, but was concentrated in Africa (97%) and pigeon pea production, which attained 4.4 million tons, was concentrated in Africa (15%) and Asia (83%). Broad beans (including *Vicia faba*, horse bean, broad bean and field bean) were also largely cultivated in 2019 with more than 5.4 million tons harvested. Lupins were produced mainly in Oceania (47%) and Europe (37%). Vetches with a production of 0.9 million tons were produced in Europe (31%) and Africa (43%) and Bambara bean was only cultivated in Africa, with an annual production of 0.2 million tons. The other pulses, grouped together for statistical purposes, accounted for 4.5 million tons worldwide. The only six pulses that are exchanged internationally are, in decreasing order of importation quantity, peas, with 37.6% of all world imports, representing 6.5 million, beans with 21%, lentils with 17%, chickpeas with 11%, broad beans with 5% and Bambara beans with only 0.01% according to the FAO data on 2016 market exchanges.

Table 1. World production of pulses in tons (T) and contribution percentage (%) of each continent (Faostat 2019).

	World (T)	Africa (%)	America (%)	Asia (%)	Europe (%)	Oceania (%)
Bambara beans	228,920	100.0	0.0	0.0	0.0	0.0
Beans, dry	28,902,672	24.4	24.4	49.7	1.3	0.3
Broad beans, horse beans, dry	5,431,503	27.0	4.0	33.6	29.4	6.0
Chick peas	14,246,295	4.9	6.1	83.4	3.6	2.0
Cow peas, dry	8,903,329	96.8	0.7	2.2	0.3	0.0
Lentils	5,734,201	3.3	42.7	42.5	2.2	9.3
Lupins	1,006,842	7.5	6.3	0.0	39.0	47.1
Peas, dry	14,184,249	3.9	39.3	18.1	37.0	1.7
Pigeon peas	4,425,969	15.1	1.8	83.2	0.0	0.0
Pulses, others	4,553,029	31.5	0.9	45.3	22.0	0.3
Vetches	762,795	42.6	12.7	12.4	31.4	0.9
Pulses, total	88,379,804	24.1	18.7	44.3	10.8	2.2

Minor pulses are produced and consumed locally worldwide, and there is no exchange between countries unless they are already cultivating and consuming these minor pulses [7]. This is more a consequence of food habits than because of pedoclimatic or agronomic constraints.

In 2018 in Europe, only 1.3% of the cultivated acreage was dedicated to pulses (Eurostat) where peas covered 37% of the surface followed by broad beans, field beans (26%) and sweet lupins (6%). The rest (26.4%) came from other dry pulses such as lentils, chickpeas and beans (Eurostat).

However, the production of pulses has decreased during the last five decades [8]. Several factors have driven this decrease. First, the reduction of human consumption of vegetal proteins was negatively correlated to the increase in the standard of living of the population and the substitution by animal proteins [9]. Besides, less time is devoted to cooking, which is essential to reduce the impact of antinutritional factors of pulses, leading to reduced consumption [10]. Second, the increasing import of cheap soybean meal to feed livestock from North and South America (2017) has induced a reduction in the price of meat. The availability of inexpensive proteins has supported the intensification of livestock production, which, at the same time, has dissociated livestock breeding from crop production for feeding [11]. The separation of protein and livestock production led to an intensification and a reduction of the number of cultivated crops, chosen for their economical performances. Moreover, as Watson et al. [8] argued, the yield increase of cereals in Europe since the 1970s was not followed by a similar yield increase of protein crops, leading to an expansion of cereal acreage while protein crops remained less attractive for farmers. However, since 2015, this trend has ended in Europe, probably because of both the increase in organic farming and the different policy instruments supporting protein crops [12].

European farmers, particularly those in arable systems, look to diversify the crop rotations to more sustainable and resilient cropping systems [13]. One of the major groups of crops targeted for diversification is grain legumes [13,14]. Several scholars have been interested in grain legumes because of their potential uses for food, feed and bioenergy/biomaterials but also because of the agroecosystem services they provide [2,3]. However, there is a general lack of knowledge and references regarding these grain legumes in terms of production potential in Northern countries and also about the diversity of their possible usage. Moreover, some scholars argue that the low adoption by European farmers of grain legumes compared with other crops is caused by low productivity and low profit margins, technological lock-in, and low temporal yield stability [15–17]. Reckling et al. [18] have underlined that in Northern Europe, the grain legume yield is as stable as that of other major spring crops, e.g., spring cereals or rape seed. Yield stability is also enhanced in the other crops of the cropping system by grain legumes, as demonstrated by Petrova Chimonyo et al. [19] in maize-based cropping systems. Mawois et al. [20] have studied the trajectories of farms that increased the proportion of grain legumes in their crop rotations, highlighting three levels for their successful introduction: the stability of the supply (both as on-farm consumption and in the food supply chain), the benefits of grain legumes as the preceding crop and/or the involvement of farmers in peer groups.

In this context, the increase in the use of grain legumes in European cropping systems can rely on two strategies. The first is to increase the current rate of cultivation of common pulses, e.g., lentils, chickpeas and common beans, in these countries, attempting to overcome the problems of lock-in at the supply chain level. This is the strategy that is currently analyzed in the mainstream literature regarding European temperate regions and in some European research and innovation projects, e.g., PROTEIN2FOOD or LEGVALUE. A complementary strategy could be to provide alternatives in the panel of grain legumes potentially cropped or processed by identifying new species from worldwide grain legume biodiversity and creating new niches for these species and a renewed interest in grain legumes. This strategy can rely, on the one hand, on the adaptability of grain legumes to new environmental conditions [21,22], and on the other hand, on the interest of the supply chain in new plant-based products, as occurred in the case of quinoa [23,24]. In this paper, our goal is to explore the latter strategy. Information on tropical grain legumes or minor pulses already exists in literature; however, it is often limited either to one species [25,26] or to a comparison of several species on a national/continental level, e.g., in India [27–29], Australia [30], Africa [31–33] or to a comparison of wild species [25,34,35]. The aim of this review is two-fold: on the one hand, it will provide insights about the agronomical and nutritional characteristics of a large panel of grain legume crops which are currently unknown and cultivated poorly or not at all in European temperate regions, and on the other hand, it will explore the potential matching of these pulses in temperate Western European climatic conditions. As we are interested in grain legumes as food, in this paper we will use the terms ‘pulses’ and ‘grain legumes’ as synonyms and distinguish them when the legume destination is not intended as food.

2. Materials and Methods

2.1. Literature Review and Data Collection

A systematic literature review was performed according to the 7 steps method formalized by the Centre for Evidence-Based Conservation [36]. The method adopted for this study is illustrated in Figure 1. The first step was the identification of the research question. We wondered about the global diversity of protein-based crops, namely of pulses, that could support a crop diversification in European temperate regions. To answer this question, we looked for a general overview of worldwide pulses diversity and characteristics. To this end, we first carried out a general literature survey in the Scopus database according to the Scopus Reference Guide [37] by selecting the advanced searches TITLE-ABS-KEY ((pulses AND ‘grain legumes’) AND crops) AND PUBYEAR > 1974 AND PUBYEAR < 2020 AND DOCTYPE (re) AND LANGUAGE (English) AND SUBJAREA

(agri) in the title, abstract and keywords with a focus on review articles, in the 1974–2019 time span in the Biological Sciences sub-database. We obtained a preliminary list of 231 reviews. Based on these reviews, we compiled a list of species to be checked. The eligibility criteria were to be cropped (not wild), used for human consumption and having a protein content in the grain equal or superior to 20 g/100 g. The species identified were double checked through a complementary search in three international databases: FoodData Central from the United States Department of Agriculture (formerly USDA National Nutrient Database for Standard Reference) [38], Prota4U hosted by Wageningen University [39] and the International Network of Food Data System from FAO [40]. A list of 47 species (common and scientific names) was finally established (Table 2). At this stage, since the information collected was too general on the agronomic and nutritional characteristics of grain legumes, in a second step we performed an in-depth literature review on the 47 previously selected species (Table 1). This second review was also performed in the Scopus database, focusing on scientific articles (therefore excluding reviews), in the 1974–2019 time span in the Biological Sciences sub-database. A grouped search including all the species was also performed in order to build a common database of minor pulses and delete duplicates. We considered minor and major pulses as separate searches.

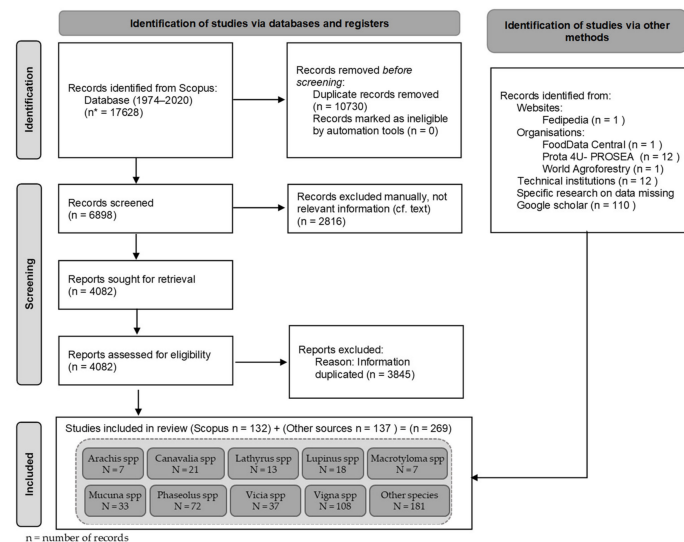


Figure 1. PRISMA 2020 flow diagram systematic review on pulses [41]. Details on the species included as well as on the number of papers retrieved are provided in Table 1.

Example of the Boolean string for this second search for one species is TITLE-ABS-KEY ((ervil OR ‘Vicia ervilia’ OR ‘Bitter vetch’) AND crop*) AND PUBYEAR > 1974 AND PUBYEAR < 2020 AND DOCTYPE (ar) AND LANGUAGE (English) AND SUBJAREA (agri) AND NOT (‘cover crop*’ OR ‘green manure’ OR intercrop*). By selecting the subject area ‘AGRI’ we tried to avoid the number of papers in ecology or natural sciences focusing on medical or beneficial properties of some of these species. Moreover, limiting the search to ‘crop’ helped us to select those papers that refer to cultivated species (not wild) and group both the common(s) and scientific names guaranteed to catch a maximum of the papers referring to a species. Finally, after a preliminary analysis of the database, we identified several papers dealing with the use of minor pulses as cover crops or green manure, along with their use in intercropping. As we were not interested in these topics, we excluded the papers including such words in the abstract, title or keywords. Even though we are aware that searching among the English literature limited the number of papers, we believed that this choice could ensure a common standard among the selected papers.

Table 2. Retrieved papers in the Scopus database for the period 1974–2019 for a sample of cultivated minor and major pulses (in grey) with a protein seed content above 20%, individual researches. In grey, a comparison with the same search was repeated for major pulses.

Common Name	Other Names	Scientific Name	Retrieved Papers in the Scopus Database (1974–2019)
Acacia leucophloea		<i>Vachellia leucophloea</i>	4
Adzuki bean	Azuki bean, aduki bean	<i>Vigna angularis</i>	102
African locust bean		<i>Parkia biglobosa</i>	31
African nut bean		<i>Ricinodeudron heudelotti</i>	1
African oil bean		<i>Pentaclethra macrophylla</i>	5
African yam bean		<i>Sphenostylis stenocarpa</i>	27
Bambara groundnut	Earth pea	<i>Vigna subterranea</i>	99
Butterfly pea		<i>Centrosema pubescens</i>	16
Cowpea	Black eye bean, black eye pea	<i>Vigna unguiculata</i>	1348
Ervil	Bitter vetch	<i>Vicia ervilia</i>	40
Fenugreek		<i>Trigonella foenum-graecum</i>	145
Grass pea		<i>Lathyrus sativus</i>	131
Guanacaste		<i>Enterolobium cyclocarpum</i>	11
Horsegram		<i>Macrotyloma uniflorum</i>	43
Housa groundnut	Hausa groundnut, Kersting groundnut	<i>Macrotyloma geocarpum</i>	6
Itching bean		<i>Mucuna pruriens var. pruriens</i>	1
Jack bean		<i>Canavalia ensiformis</i>	36
Kedaung	Tree bean	<i>Parkia roxburghii</i>	2
Kidney bean		<i>Phaseolus vulgaris</i>	131
Lablab	Hyacinth bean	<i>Lablab purpureus</i>	125
Lima bean	Butter bean, Java bean, Madagascar bean, sugar bean	<i>Phaseolus lunatus</i>	113
Marama bean	Morama bean	<i>Tylosema esculentum</i>	8
Moth bean	Mat bean, Turkish gram	<i>Vigna aconitifolia</i>	63
Mung bean	Golden gram, green gram, Moong bean	<i>Vigna radiata</i>	811
Navy bean	Pearl haricot, haricot bean	<i>Phaseolus vulgaris</i>	66
Narbon bean	Narbon vetch	<i>Vicia Narbonensis</i>	43
Pigeon pea	Red gram	<i>Cajanus cajan</i>	590
Pinto bean		<i>Phaseolus vulgaris</i>	58
Pinto peanut		<i>Arachis pintoi</i>	70
Purple mucuna		<i>Mucuna atropurpurea</i>	0
Red moneywort		<i>Alysicarpus rugosus</i>	1
Rice bean		<i>Vigna umbellata</i>	55
Scarlet runner bean	Runner bean	<i>Phaseolus coccineus</i>	18
Sesban		<i>Sesbania sesban</i>	46
Sword bean		<i>Canavalia gladiata</i>	9
Tamarind		<i>Tamarindus indica</i>	45
Tarwi		<i>Lupinus mutabilis</i>	30
Tepary bean		<i>Phaseolus acutifolius</i>	46
Urad	Black bean, black lentil, mungo bean	<i>Vigna mungo</i>	357
Velvet bean	Cowitch	<i>Mucuna pruriens var. utilis</i>	25
Winged bean		<i>Psophocarpus tetragonolobus</i>	39
Yam bean		<i>Pachyrhizus erosus</i>	45
Broad bean	Faba bean, horse bean	<i>Vicia faba</i>	918
Chickpea	Bengal gram	<i>Cicer arietinum</i>	1795
Common bean	Field bean, bell bean, English bean, Windsor bean, pigeon bean	<i>Phaseolus vulgaris</i>	1988
Field pea		<i>Pisum sativum</i>	2014
Lentil		<i>Lens culinaris</i>	901
Lupin		<i>Lupinus spp.</i>	688

Then, a second search was performed in Google Scholar according to the same criteria as in the Scopus database in the case of absent or insufficient number of papers (e.g., Purple mucuna or Red moneywort), or in cases where the papers retrieved in the Scopus database did not target our goal, e.g., focusing exclusively on genetics, crop protection or agroforestry potential or looking only for a specific characteristic in growth, agronomy, nutritional or antinutritional composition. For minor pulses with a limited or local use, nutritional characteristics were obtained from research articles, of which a mean was calculated (protein, oil, carbohydrates and fiber content) or an interval was indicated.

The papers obtained from the Scopus searches for each pulse species were stored in a Zotero database, which contained a total of 17,628 articles, and 10,730 of them were discarded as duplicates. Among the duplicates, there were often papers comparing two or several pulses. An exhaustive screening on titles was performed in order to eliminate out-of-scope papers that did not provide useful information for our research (6661 papers). The most common reasons for deleting a paper from the list were the following: the name of a pulse was mentioned only as a comparison of one characteristic to another crop [42], the trial or analysis was performed in intercropping, thus evaluating the impact of the cover crop on the staple food yield [43] or there was no relation to the topic, e.g., papers dealing with biochemical [44] or genetic characteristics [45] or the use of pulse extracts as green fertilizers [46] or seed treatments [47] without any relation to pulse yield or seed quality. Papers with an historical or archeobotanical approach [48] and those stating consumer preferences [49] or the impact of the feed on the animal's health [50] were also excluded. For some crops, such as cowpea or Bambara bean, not all the articles found are cited in the present review, as for one topic, such as resistance to a fungus or an insect, dozens of articles could be found. Some cultivated legumes showed an interesting seed composition but they are not used as food. Brebra seed (*Millettia ferruginea*) was excluded from our analyses, even though the seed is edible. Other species from *Lathyrus* and *Vicia* with protein content above 20% were used as food but are either no longer cultivated [51] or are cultivated on a few hectares of a single Greek island for traditional dishes like *Lathyrus clymenum* [52]. Finally, papers dealing with consumer preferences for different varieties or for different cooking techniques were also excluded.

In the end, the final number of papers used for the review was 269.

2.2. Bibliographic Corpus Analysis

The file containing the Scopus extraction with information from 4082 documents assessed for eligibility was first analyzed to describe the papers in terms of sources (journals), years of publication and origin. For these analyses, we calculated the absolute, relative and cumulative frequencies. Then, the corpus was uploaded and analyzed using the CorTexT platform, already used for agronomic literature reviews [53,54], through-out the CorTexT Manager (INRAE-LISIS, Noisy-le-Grand, Seine-Saint Denis, France) (<http://manager.cortext.net/>) (accessed on 30 November 2021).

A term-extraction algorithm was first performed using the title, abstract and keywords of each document to identify the 100 most cited terms. With those terms, several functions have been tested in the default proposed form such as the contingency matrix, geospatial exploration and network mapping to see the relationships between species, countries, journals and other agronomical terms.

2.3. Descriptive Analysis

The aim of the descriptive analysis was to characterize both the agronomic and nutritional characteristics of the selected species. Both are essential to select candidate species diversifying cropping systems in temperate regions to be used in the food industry. On the one hand, for the agronomic characteristics, we analyzed the environmental requirements (optimal, minimal and maximal temperature, average water requirement, duration of crop cycle), fertilizer inputs, potential yield, fixed nitrogen and symbiotic bacteria. On the other hand, for the nutritional characteristics, we analyzed the nutritional profile (calories,

protein content, carbohydrates, vitamins, minerals, micronutrients) and the most common antinutritional characteristics (phytic acid, total tannins, saponins, trypsin inhibitors, lectins and other antinutritional molecules).

2.4. Species Selection for Temperate Regions

The most promising crops for arable systems in temperate regions were selected from the database mainly according to the environmental requirements. Focusing on arable systems, only annual crops that could rotate with other crops were considered. For a preliminary evaluation, we also excluded crops that require special harvesting techniques such as those that produce grains underground. For annual species we considered those having varieties with maximum crop duration of 120 days, thus corresponding to summer crops in temperate regions. We also excluded species with optimal temperatures higher than 25 °C or where those data were absent. We hypothesize for the annual crops a growing period from 1 March to 15 October. We could not consider the crop heat units to reach maturity, the sensitivity to day length and the precocity because of a lack of data in the literature. We considered as candidate species those that can potentially grow in continental European temperate regions under oceanic influence (by excluding temperate areas in Ireland) according to the European environmental stratification proposed by Metzger et al. [55]. Climatic data (1998–2018) were extracted on the Agri4cast database provided by the European Joint Research Centre (<https://agri4cast.jrc.ec.europa.eu/DataPortal/Index.aspx>) (accessed on 11 January 2021) in the area between latitudes 47.56° to 53.44° and longitudes −0.017° to 7.28°. The spring sowing date (date to start accumulating growing degree days (GDD)) was considered as the last of three consecutive days with daily mean air temperatures equal to or greater than 12.8 °C based on the Corn Heat Units Index used to characterize climatic regions [56]. The potential season-ending date (date to stop accumulating GDD) was considered as the earliest date when the daily mean air temperature dropped to 12 °C, excluding dates included in the 90 days following the sowing date. The sum of temperatures was calculated within those dates considering base temperatures of 6 °C and 10 °C. Given the lack of data on the grain legumes investigated, those climatic data were compared to the requirements of already cultivated spring crops such as corn or soybean, to establish preliminary conformity of the criteria.

3. Results and Discussions

3.1. Descriptive Analysis of the Bibliographic Corpus

The selected corpus on pulses was composed of 4081 documents. More than 50% of the research was published since 2010 (data not shown). The observed bibliography shows that the interest in pulses has been increasing, especially in the last 15 years, reaching more than 250 documents per year (data not shown). An analysis of the country of origin of the first authors shows that affiliations from 116 countries are represented; of them, more than 50% are from two countries, Brazil and India. Papers from European temperate areas are poorly represented. Finally, more than 600 individual journals are represented in the corpus, and 55 account for 50% of the articles. Of these latter journals, mostly are in the agronomy and crop science area, e.g., *Crop Science*, *Field Crop Research* or *Experimental Agriculture*, and have partially regional coverage, e.g., *Indian Journal of Agricultural Sciences* or the *Australian Journal of Crop Science* (data not shown). The matrix of contingency obtained from the CorTexT platform (data not shown) also confirmed that almost 40% of the retrieved papers were published in two Indian journals (*Indian Journal of Agricultural Sciences* and *Indian Journal of Agronomy*) and more than 40% on *Phaseolus vulgaris* (or common bean, data not shown). This was confirmed by geospatial exploration which identified India as the main producer of references on pulses, ahead of other countries in the world.

The most cited terms were *Phaseolus vulgaris* (in different written forms) with 537 occurrences which corresponded to more than 25% of all the literature, followed by *Vigna radiata* 407, *Cajanus cajan* 238, *Experimental field* 155, *Triticum aestivum* 152, *Vigna mungo* 142, *Common bean* 95, *Pisum sativum* 70 and *Arachis hypogaea* 69. This confirms

as annuals depending on the agronomical practices of the country. Despite the in-depth research conducted it was difficult to identify the intra species genetic diversity available that may be conducive to a larger potential for one species to spread to other climatic conditions or agronomical practices.

In temperate climates, pulses are mainly cultivated as main crops or in association with cereals [65], whereas in tropical countries, we can find associations with corn [66], coffee [67], cassava [68] and other diverse crops [69,70]. In the latter crop, little or no fertilization or pest management is required. Two pulses are cultivated not for their grains but for their tuber production, making the grains a coproduct (yam bean and African yam bean [71,72]). Among the studied species, minimal temperature has been retrieved in 25 species over 57. Among them, a few minor pulses resist cold temperatures (Tarwi bean, pigeon pea, kidney bean and fenugreek). Among the major pulses, five over six tolerate negative temperatures. The other pulses require positive temperatures during their entire life cycle, and in most cases, above 10 °C. The optimal growing temperature is characterized in the literature only in 33 out of 47 species, but the temperature intervals are often wide. The maximal temperature is retrieved in 23 species over 47 but it will not be a constraint in temperate climates of Western Europe.

In temperate climates, pulses are mainly cultivated as main crops or in association with cereals [65], whereas in tropical countries, we can find associations with corn [66], coffee [67], cassava [68] and other diverse crops [69,70]. In the latter crop, little or no fertilization or pest management is required. Two pulses are cultivated not for their grains but for their tuber production, making the grains a coproduct (yam bean and African yam bean [71,72]). Among the studied species, minimal temperature has been retrieved in 25 species over 57. Among them, a few minor pulses resist cold temperatures (Tarwi bean, pigeon pea, kidney bean and fenugreek). Among the major pulses, five over six tolerate negative temperatures. The other pulses require positive temperatures during their entire life cycle, and in most cases, above 10 °C. The optimal growing temperature is characterized in the literature only in 33 out of 47 species, but the temperature intervals are often wide. The maximal temperature is retrieved in 23 species over 47 but it will not be a constraint in temperate climates of Western Europe.

3.3. Nutritional Characteristics

Pulses have an interesting nutritional profile (Table 4, Appendix B), because they have high protein content with a mean value in our sample of 23% (rice bean has the lowest content with 19.7% [75] and tarwi [40] has the highest content, 51%), low fat content with a mean value of 3.8% and a small percentage of saturated fatty acids. Pulses also provide fiber, vitamins and minerals which are important for human health. Table 4 and Appendix B show the nutritional characteristics of pulses, not taking into account genetic or pedoclimatic variability between species. Vitamins and minerals, when present, were included in Appendix B. It is interesting to note that the values of protein content in minor pulses were comparable to or higher than those of the common pulses already cultivated in North-West Europe.

Table 3. Pulses' agronomic requirements and potential yields, extracted from Appendix A.—indicates that no data were retrieved from the literature.

Pulses—Common Name	Temperature (°C) Min; Max; Optimal	Crop Length	Water Requirements (mm)	Yield (t/Ha)	Kg N Fixed (kg/ha)
Adzuki bean	5–10; 34; 15–30	60–190 d	500–1700	0.5–3.5	100
Butterfly pea	5; 35; 20–28	120–194 d	400–1750	0.7	-
Cowpea	15; 35; 25–35	60–340 d	500–1500	0.2–7	12–50
Fenugreek	–4; -; 18–27	90–100 d	300–400	-	-
Grass pea	20; -; 10–25	90–180 d	400–650	0.3–10.5	25–50
Horse gram	-; 40; 20–32	120–150 d	380–900	0.5	-
Itching bean	-; -; 19–27	-	400–3000	0.2–2	-
Kidney bean	–1; 30; 15–20	65–105 d	300–600	2.2	20–44
Lablab	-; -; 18–30	54–220 d	650–3000	1.4–4.5	20–140
Lima bean	>0; >37; 16–27	115–180 d	900–1500	0.4–5	40–60
Moth bean	25; 45; 24–32	70–90 d	200–750	0.07–2.6	-
Navy bean	12; 35; 22–30	53–300 d	400	0.5–5	125
Pigeon pea	0; 40; 18–29	3–5 y	600–1400	0.6–5	69–134
Pinto bean	-; 36; 21–25	90–100 d	-	0.5–3.9	-
Rice bean	10; 40; 25–35	120–150 d	700–1700	0.2–2.7	-
Sword bean	-; -; 25–30	150–300 d	900–1500	1.5–5.4	75–230
Tarwi	<0; -; -	150–330 d	-	0.6–4.8	100
Tepary bean	8; >32; 17–25	60–120 d	400–1700	0.4–1.7	-
Urad	-; -; 25–35 no frost	60–140 d	600–1000	0.3–2.5	18
Velvet bean	5; -; 19–27	-	1200–1500	0.5–3.4	60–330
Winged bean	-; -; 20–30	120–180 d	1000	0.7–1.9	-
Broad bean	–12; 30; 18–27	90–220 d	700–1200	1.1–2.2	33–550
Chickpea	–11; >32; 10–29	90–180 d	500–1800	1–5.5	35–140
Common bean	–9; 38; -	70–110 d	274–550	0.9–2.6	465
Field pea	<0; -; 7–30	90–180 d	400–1000	1–4	30–96
Lentil	2; 35; 6–27	80–130 d	300–2400	0.8–7	50
Lupin	<0; -; 18–24	115–330 d	400–1000	0.5–5	90–400

d means days (annual crops) and y years (pluriannual crops), °C degrees Celsius. In grey, a comparison with the same search was repeated for major pulses.

Other current and largely cultivated crops are also rich in proteins (Appendix C) but often they also have high fat content, making them less nutritionally valuable. Appendix C describes in the same way as Appendix B the nutritional characteristics of other protein-rich crops cultivated or not in temperate regions. For cotton and rapeseed, there are nearly no data concerning their nutritional composition as they are mainly used to obtain fiber and oil or oil and proteins, respectively, with no direct use as food or feed. A common protein supplement, spirulina (*Arthrospira maxima*) was also included because of its interesting nutritional profile, highest content of proteins and third lowest level of oil content by comparison.

Table 4. Nutritional profile of pulses, extracted from Appendix B.

Pulses— Common Name	Energy (kcal/100 g)	Protein (%)	Oil (Saturated FA) (%)	Carbohydrate (%)	Fiber (%)
Adzuki bean	329	19.9	0.5 (0.2)	62.9	12.7
Butterfly pea	-	25.2	3.7	19.9	9.2
Cowpea	343	23.9	2.1 (0.5)	59.6	10.7
Fenugreek	323	23.0	6.4	58.0	25.0
Grass pea	-	24.4	2.8 (0.8)	55.94	11.4
Horse gram	280	22	0.6	37.5	5.7
Itching bean	382	27–37	6.6–8.8	46–53	6–10
Kidney bean	333	23.6	0.8 (0.1)	30.0	24.0
Lablab	344	21–29	1.7 (0.3)	60.74	25.6
Lima bean	338	21.5	0.7 (0.2)	63.4	19.0
Moth bean	343	23–26	1.6 (0.4)	61.5	5
Navy bean	337	22.3	1.5 (0.2)	60.8	15.3
Pigeon pea	343	13–26	1.5 (0.3)	62.8	15
Pinto bean	347	21.4	1.2 (0.2)	62.6	15.5
Rice bean	338	18–19	0.5 (0.3)	59.1	7.1
Sword bean	361	24–30	2.6–9.8 (-)	41–59	7–13
Tarwi	440	41–51	14–24 (19)	28.2	7.1
Tepary bean	353	19–24	1.2 (-)	67.8	4.8
Urad	341	25.2	1.6 (0.1)	59.0	18.3
Velvet bean	373	20–29	6–7	50–61	9–11
Winged bean	428	30–35	16.3 (2.3)	41.7	11–26
Broad bean	341	26.1	1.53 (0.3)	58.3	25
Chickpea	378	11–31	6.0 (0.6)	63.0	12.2
Common bean		20–24	0.8 (0.6)	75.5	-
Field pea	352	23.8	1.2 (0.2)	63.7	25
Lentil	352	24.6	1.1 (0.2)	63.4	10.7
Lupin	371	36.2	9.7 (1.2)	40.37	18.9

In grey, a comparison with the same search was repeated for major pulses.

3.4. Antinutritional Characteristics

All pulses contain one or more antinutritional compounds (Appendix D) and although the concentration is under the lethal dose, a regular or almost exclusive consumption may induce certain medical problems, such as the lathyrism caused by grass pea in European populations after the wars [76] that led to a law forbidding the consumption of grass pea grain or its derivatives in Spain [77]. Efforts have been made to breed new varieties with reduced content of the neurotoxin [78].

Appendix D reports data from research articles that analyzed one or several compounds. The other columns indicate the presence of other toxic molecules that are only present in some crops, even in a single crop, such as β -ODAP (beta-oxalyl-diaminopropionic acid) in grass pea or gossypol in cotton. Itching and velvet bean present the highest content of phytic acid, with some varieties reaching between 53 and 57 mg/g [28], whereas the other pulses varied from 0.5 to 41 mg/g. In the case of tannins, some varieties of African yam bean contained up to 18.1 mg/g [79], while other pulses contained from 0.01 [80] to 96 [81] mg/g. Saponins reached the highest values in some varieties of navy bean, but have lower values than some varieties of lentils or chickpea [82]. Trypsin in-

hibitors were found in high quantities in sesban seeds, reaching 140 mg/100 g [83]. Finally, lectins were less abundant in a few species of pulses; mung bean seeds contained the highest quantity, with 15.8 mg per 100 g [84].

No pulse exceeded the lethal dose estimated at 50–60 mg/kg for phytates, 30 mg/kg for tannins, 2.5 g/kg for trypsin inhibitors, 50 mg/kg for lectins, 50–60 g/kg for hydrogen cyanide or 20 mg/100 g and 2–5 g/kg for oxalates [85]. Nevertheless, the presence of these antinutritional compounds may cause some troubles. Phytic acid chelates with several minerals, limiting their bioavailability for the organism. Tannins reduce the absorption of nutriment and vitamin B12, reducing the efficiency of energy conversion [86]. Cooking or transforming these pulses inactivates the negative effects of these antinutritional factors on health or wellbeing [87].

3.5. Potential for Minor Pulse Cultivation in European Temperate Regions

Analyses of the meteorological data showed that in the considered European temperate regions between 1998 and 2018, the total number of observed situations was 7456. The minimal duration of good cropping conditions was 121 days, and the average maximum was 140 days. The most frequent duration was 120–125 days (18.5%) followed by 125–130 (15.3%) and 130–135 days (10.5%). The daily temperature varied from 14.9 to 20.0 °C, with a mean of 17.1 °C. Sowing was possible in 12.6% of the situations in March, 54.1% in April, 34.0% in May and only 1.8% in June, whereas the possible harvest day was concentrated between September (64.7%) and October (33.6%). The GDD in base 6 °C varied from 1102 to 2430 °C day, the most frequent being 1500 to 1600 °C day (19%), 1600–1700 °C day (18.3%), 1400–1500 (16.2%) and 1700–1800 (13.2%). The GDD in base 10 °C varied from 615 to 1654 °C day with a mean of 994 °C day being the most frequent, 1000–1100 (25.5%), followed by 1100–1200 (20.6%) and 900–1000 (20.2%) (Figure 3).

By applying the criteria of crop needs and the possibility of reaching the GDD necessary to attain crop maturity, compared to other spring crops already cultivated in those regions, almost half the selected grain legume crops (21) appeared to be of interest for temperate cropping systems. Those species are the ones listed in Tables 2 and 3.

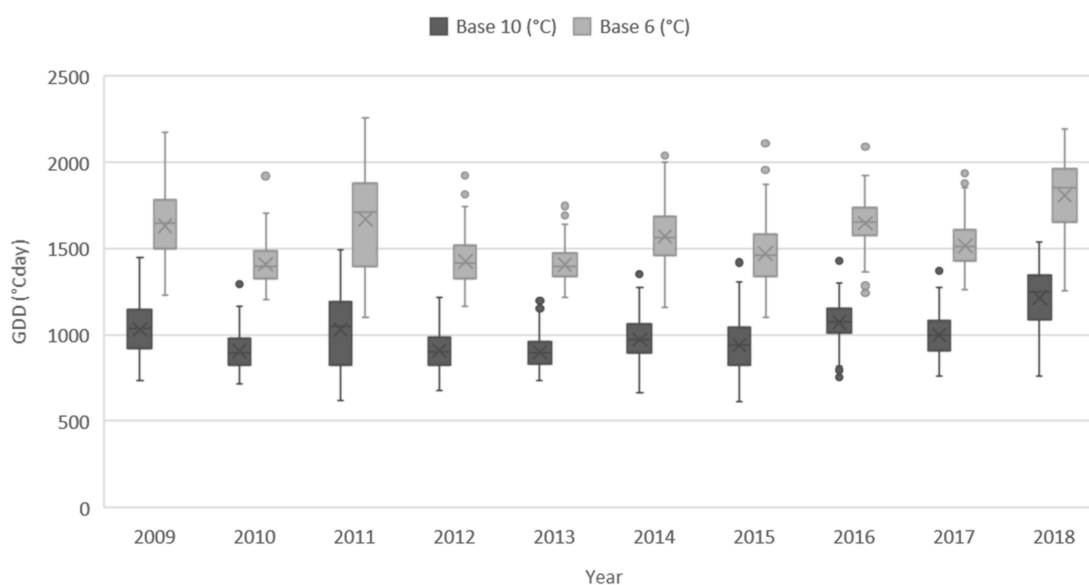


Figure 3. Bow whiskers of the growing degrees days (GDD, °C day) cumulative distributions in several locations in European temperate regions (see Section 2.4) within the last 10 studied years for a base temperature of 6 and 10 °C (source: own calculation from the Agri4cast database).

4. Conclusions

The present systematic review has revealed the agronomic and nutritional interest of almost 20 pulses beyond the major ones which could be cultivated in Western European temperate regions. At this stage, this possibility is theoretical as there are several gaps in the literature. Firstly, there is a lack of physiological data, especially for minor pulses, whereas for major pulses information could be retrieved but only for the current cultivation area. Particularly, data concerning the optimal and minimal growing temperatures, the GDD requirements to reach physiological maturity as well as the minimal water requirements are missing. Secondly, there is little information about soil constraints such as the soil structure, soil depth, pH or soil water availability. Pests and diseases are described for most pulses, even for those species with few references. Thirdly, beyond the species level, there is a lack of knowledge about the genetic diversity that could be useful to extend the cultivation area, as the characterization that is retrieved in scientific literature is not sufficient to the best of our knowledge. Probably, in seedbanks, the information available will be more exhaustive for assessing certain traits such as crop length or disease resistance but this information will not be enough to establish the potential crop length in climatic areas other than those where genetic material has been collected. Among the 20 pulses with the highest cultivation potential in temperate areas, we have found that both their nutritional and antinutritional characteristics are comparable to those of the major pulses already cropped and consumed in these areas. Technological characteristics, processing potential and consumer preferences were outside of the scope of this review; however, all the retrieved species are already consumed as raw food or food components in different parts of the world. The potential of these species for uses other than as food, i.e., feed (grain or forage) or non-food (bioplastic) were also outside of the scope of this literature review but they can be considered worthy of investigation when contemplating the development of a new supply chain.

From these conclusions, we can highlight three main research perspectives when addressing the introduction of these pulses in Western European temperate areas. These are traditional perspectives when considering the introduction of a new crop to different environments, as occurred for example with quinoa. Firstly, a characterization of the requirements of the pulses in a controlled environment is necessary to identify their growing needs in order to adapt the crop management to obtain the maximum potential yield. Then, multisite and pluriannual field trials are required to establish the optimal pedoclimatic conditions and management practices. In parallel, a second perspective concerns the great effort that must be undertaken to study the existing genetic diversity in order to find the varieties that are most adapted to temperate conditions. This effort will require a large collaboration with seedbanks in order to characterize this diversity. Finally, a third perspective will concern a breeding program to improve the existing varieties. These three perspectives are a necessary first step before considering adoption by local farmers and introduction to cropping systems in temperate areas.

Author Contributions: Conceptualization, A.A.G. and E.M.; methodology, A.A.G. and E.M.; formal analysis, A.A.G. and E.M.; writing—original draft, A.A.G. and E.M.; writing—review and editing, A.A.G. and E.M.; supervision, A.A.G. and E.M. All authors have read and agreed to the published version of the manuscript.

Funding: This review was partially funded by the SFR Condorcet with the project CALEGE 2021.

Data Availability Statement: The datasets analyzed during the current study are available from the corresponding author on reasonable request.

Acknowledgments: We acknowledge the contribution in the first stage of the literature review of Sophie Bonningues and Gauranvi, interns at UniLaSalle (France). We also acknowledge our colleagues Michel-Pierre Faucon and Pierre Saulet for their useful discussions and the Agri4cast data extraction.

Conflicts of Interest: The authors declare no conflict of interest.

Appendix A

Table A1. Agronomic requirements of minor and major pulses in grey major pulses.

Pulses—Common Name	Climate Region	Countries Where Cultivated	Temperature (°C) Min; Max; Optimal	Crop Length ^a	Water Requirements (mm)	Mineral Re-quirements	Pest Risk ^b	Yield (t/ha)	Kg N Fixed (kg/ha)	Bacteria Associated	References
Acacia leucophloea	Tropic	Bangladesh, India, Indonesia, Pakistan	6; 49; -	Y (tree)	400–1500		F; I				[88]
Adzuki bean	Tropic	China, Japan, Korea, Nepal	5–10; 34; 15–30	60–190 d	500–1700	N, P, K	B; F; I	0.5–3.5	100	<i>Bradyrhizobium</i> spp.; <i>Sinorhizobium fredii</i>	[89–97]
African locust bean	Tropic	Sudan, Uganda	0; 45; 21–36	Y (tree)	500–1500		B; F; I				[98,99]
African nut tree	Tropic	West and Central Africa, Madagascar	-; -; 18–32	Y (tree)	1000						[39,99]
African oil bean	Tropic	Sub-Saharan Africa	18; -; 25	Y (tree)	1000–2000		I				[39,99]
African yam bean	Tropic	Ghana, Nigeria, Togo	0; -; 19–27	150–240 d	900–2000		F; I	3 A *			[90,98,100]
Bambara groundnut	Tropic	Australia, Cameroon, Nigeria, Sudan	15; 40; 20–28: no frost	90–180 d	600–1200	N, P, K	F; I; N; V	0.1–6	100	<i>Bradyrhizobium</i> spp.	[90,101–109]
Butterfly pea	Tropic	Brazil, Colombia, India, Mexico, Philippines, Venezuela	5; 35; 20–28	120–194 d	400–1750		F; I; B	0.7		<i>Rhizobium</i>	[110–114]
Cowpea	Tropic	Burkina Faso, Ghana, Mali, Senegal	15; 35; 25–35 no frost	60–340 d	500–1500	N, P, K	F; I; P; PW; V	0.2–7	12–50	<i>Rhizobium</i> spp. NGR234; <i>Bradyrhizobium</i> spp.; <i>Sinorhizobium fredii</i>	[39,62,100, 115–124]
Ervil	Temperate	Australia, Iran, Morocco, Turkey		134–154 d	200–500			0.6–4			[125–129]
Fenugreek	Temperate	Morocco, Tunisia, Spain	-4; -; 18–27	90–100 d	300–400	N, P, S				<i>Rhizobium</i> spp. NGR234	[130,131]
Grass pea	Temperate/subtropic	India, Iran, Italy, Middle East, Spain	20; -; 10–25	90–180 d	400–650	N, P	F; I; V	0.3–105	25–50		[39,63,98,132, 133]
Guanacaste	Tropic	Brazil, Costa Rica, Mexico, Venezuela	-; -; 23–28	Y (tree)	750–2000						[134]
Horse gram	Tropic	Australia, India, USA	-; 40; 20–32: no frost	120–150 d	380–900	N, P	F; I	0.5		<i>Bradyrhizobium</i>	[39,90,100,118, 135,136]
Housa groundnut	Tropic	Cameroon, Nigeria, Senegal	18; 34; 25–32	70–180 d	500–600	Few	F	0.5	42	<i>Bradyrhizobium</i> <i>Rhizobium</i> spp.	[39,137,138]
Itching bean	Tropic	India	-; -; 19–27		400–3000			0.2–2			[139]
Jack bean	Tropic	Brazil	-; -; 13–27	180–300 d	800–2000		N	4.5			[100,140,141]
Kedaung	Tropic	Bangladesh, India, Egypt, Malaysia				N, P, K	F, I				[142]
Kidney bean	Temperate	Brazil, China, EU, India, Mexico, USA	-1; 30;15–20	65–105 d	300–600	N, P, K, S, Zn	F; I; N; V	2.2	20–44	<i>Rhizobium</i>	[98,130,132, 143]
Lablab	Tropic	Cameroon, India, Madagascar	-; -;18–30	54–220 d	650–3000	Few	F; I; N; V	1.4–4.5	20–140	<i>Rhizobium</i> spp. NGR234	[63,90,98,116, 144–146]
Lima bean	Temperate	Brazil, Mexico, Peru	>0; >37; 16–27	115–180 d	900–1500	N, P, K, Zn	B; F; I; N; V	0.4–5	40–60	<i>Bradyrhizobium</i> <i>Rhizobium</i> spp.	[39,90,98,147–149]
Morama bean	Tropic	India	-; 37; -	Y	100–900						[39,137]
Moth bean	Tropic	Australia, India, Pakistan, Thailand, USA	25; 45; 24–32	70–90 d	200–750		I; N; V	0.07–2.6		<i>Bradyrhizobium</i> <i>Rhizobium</i> spp. NGR234	[39,116,150–152]
Mung bean	Temperate	China, India	12; 40; 28–30	50–120 d	600–1000	P, K, Ca, Mg, Zn	B; F; I; V	0.3–2.2	14	<i>Rhizobium inoculums</i>	[39,92,98,102, 153–156]
Navy bean	Temperate/Tropic	Australia, EU, USA	12; 35; 22–30	53–300 d	400	N, P, Zn	B; F; I	0.5–5	125	<i>Rhizobium phaseoli</i>	[39,130,157, 158]
Narbon bean	Temperate	Australia, Iraq, Italy, Jordan, Portugal, Spain, Turkey	30; -; -	170 d	200–500		PW; F; I; N; V	0.5–2			[30,126,127, 159–161]
Pigeon Pea	Tropic	India, Kenya, Malawi, Myanmar, Nepal, Tanzania, Uganda	0; 40; 18–29: no frost	3–5 y	600–1400	P	I; N	0.6–5	69–134	<i>Bradyrhizobium</i> spp.	[63,92,98,102, 141,162–164]
Pinto bean	Temperate	USA	-; 36; 21–25	90–100			B; V	0.5–3.9			[165,166]
Pinto peanut	Tropic	Argentina, Brazil, Colombia, USA	-; -; 21–30		1100–1500	K, Al, Mn	F; N; V	0.3–3			[63,167–169]
Purple muncuna	Tropic	India	-; 19–27; -	Y	400–3000						
Red moneywort	Temperate/Tropic	Australia, Asia, Madagascar			600–1500			3–7.5		<i>Bradyrhizobium</i>	[170]
Rice bean	Temperate-Tropic (Alt≈2000 m)	Bangladesh, China, India, Nepal	10; 40; 25–35	120–150 d	700–1700	P	B; F; I	0.2–2.7		<i>Rhizobium</i> spp. NGR234	[39,90,102,118, 171–174]

Table A1. *Cont.*

Pulses—Common Name	Climate Region	Countries Where Cultivated	Temperature (°C) Min; Max; Optimal	Crop Length ^a	Water Requirements (mm)	Mineral Re-quire-ments	Pest Risk ^b	Yield (t/ha)	Kg N Fixed (kg/ha)	Bacteria Associated	References
Scarlet runner bean	Temperate-Tropic (Alt≈2000 m)	Africa, Central & South America, EU	5; >37; 25	120–150 d	1500	F	F;	0.9–12.5		<i>Rhizobium</i>	[39,149,175]
Sesban	Tropic	Chad, Egypt, Kenya, Uganda	7; 45; 17–20	Y (tree)	500–2000	P, K	F; I; N; V			<i>Rhizobium leguminosarum</i> , <i>Bradyrhizobium</i>	[98,170,176–178]
Sword bean	Tropic	India	-; -; 25–30	150–300 d	900–1500		F; I; N	1.5–5.4	75–230		[39,98]
Tamarind	Tropic	Australia, Cameroon, China, India, Mexico, Nigeria	4; 41; 15–28	Y (tree)	32–3800	P	B, F; I; N				[179–183]
Tarwi	Temperate (Alt≈3000 m)	South America	<0; -; -	150–330 d		N, P	F; I; V	0.6–4.8	100		[90,184]
Tepary bean	Temperate/Tropic	Guatemala, Mexico, USA	8; >32; 17–25	60–120 d	400–1700	P	B; F; I; N; V	0.4–1.7		<i>Bradyrhizobium</i> , <i>Rhizobium leguminosarum</i> <i>bc. phaseoli</i>	[39,90,130,149,185]
Urad	Tropic	India, Pakistan	-; -; 25–35 no frost	60–140 d	600–1000	P, B	F; I	0.3–2.5	18	<i>Bradyrhizobium yuanmingense</i>	[62,153,154,186]
Velvet bean	Tropic	India	5; -;19–27	D or Y	1200–1500		I; N	0.5–3.4	60–330	<i>Bradyrhizobium</i> sp.	[90,98,187–191]
Winged bean	Tropic	India, Indonesia, Philippines, Sri Lanka	-; -;20–30	120–180 d	1000		F; I	0.7–1.9		<i>Rhizobium</i> spp. NGR234	[116,192]
Yam bean	Tropic	Costa Rica, India, Mexico, Peru	-; 35–40; -	2 Y		N, P, K	N; I	A *	165–215	<i>Rhizobium</i> spp. NGR234	[64,116,193,194]
Broad bean	Temperate/Tropic	Australia, China, Ethiopia, EU, Jordan, USA	-12; 30;18–27	90–220 d	700–1200	N, P, K, Ca	F; I; N; V	1.1–2.2	33–550	<i>Rhizobium Leguminosarum</i>	[39,92,98,130,132,195,196]
Chickpea	Temperate/Tropic	EU, Middle East, South Africa	-11; >32; 10–29	90–180 d	500–1800	P, Zn, S, B	F; I; N; V	1–5.5	35–140	<i>Rhizobium cicerii</i>	[39,92,98,130,153,195,197–201]
Common bean	Temperate	Australia, Africa, Canada, EU, Middle East, USA	-9; 38; -	70–110 d	274–550	N, P, S, Mo	F; I; V	0.9–2.6	465	<i>Rhizobium leguminosarum</i> ; <i>Rhizobium tropici</i>	[202–213]
Field pea	Temperate/Tropic	Canada, China, EU, India, Russia	<0; -; 7–30	90–180 d	400–1000	N, P, K, Mg	F; I; N; V	1–4	30–96	<i>Rhizobium leguminosarum</i>	[39,92,98,130,132,156,195,214]
Lentil	Temperate	Australia, Canada, EU, Middle East	2; 35; 6–27	80–130 d	300–2400	N, P, K	F; I; N; V	0.8–7	50	<i>Rhizobium</i>	[39,92,98,102,130,153,195,215,216]
Lupin	Temperate	Australia, EU, Middle East, Ukraine	<0; -;18–24	115–330 d	400–1000	P, Fe	F; I; V	0.5–5	90–400	<i>Rhizobium lupini</i>	[39,92,98,132,195,217–220]

^a Crop length: days (d), years (Y); ^b pests: fungi (F), insects (I), bacteria (B), nematodes (N), virus (V), parasite weeds (PW); * A: the tuber of the plant is the main crop use.

Appendix B

Table A2. Nutritional composition of minor pulses, in grey major pulses.

Pulses—Common Name	Energy (kcal/100 g)	Protein (%)	Oil (Saturated FA) (%)	Carbohydrate (%)	Fiber (%)	Vitamins	Minerals	Reference
Acacia leucophloea	382	27	5	58	7		Ca, Fe, Mg, P, K, Na, Zn, Cu, Mn	[88,221]
Adzuki bean	329	19.9	0.5 (0.2)	62.9	12.7	B ₁ , B ₂ , B ₃ , B ₆ , B ₉ , A	Ca, Fe, Mg, P, K, Na, Zn	[38,222]
African locust bean	414	24–34	19–23	67	11.7	C	Ca, Fe, Mg, K, Na, Zn, Cu, Mn	[99,223–227]
African nut bean	649	26.3	58.1	4.6	2.7		Ca, Fe, Mg, P, Zn, Cu	[99]
African oil bean	206	48.5	33.4	8.9	6.3	C, B ₁ , B ₂ , B ₃	Ca, Fe, Mg, P, K, Na, Cu, Mn, Pb	[71,99,226,228]
African yam bean	365	20.5	1–12.2	65–78	7–12		Ca, Fe, Mg, P, K, Na, Zn, Cu, Mn	[71,79,229,230]
Bambara groundnut	408	18–30	6.2 (2.0)	33.4–68.5	1.9	C, B ₁ , B ₂ , B ₃ ,	Ca, Fe, Mg, P, K, Na, Se	[40,104,231–235]

Table A2. Cont.

Pulses—Common Name	Energy (kcal/100 g)	Protein (%)	Oil (Saturated FA) (%)	Carbohydrate (%)	Fiber (%)	Vitamins	Minerals	Reference
Butterfly pea	-	25.2	3.7	19.9	9.2		Ca, P	[236]
Cowpea	343	23.9	2.1 (0.5)	59.6	10.7	C, B ₁ , B ₂ , B ₃ , B ₆ , B ₉ , A	Fe, Mg, Zn, Cu, Mn, Cr, Ni, Al, Pb	[38,40,237,238]
Ervil	324	20–28	11–16	61			Ca, Fe, Mg, P, K, Na, Zn, Cu, Mn	[239–243]
Fenugreek	323	23.0	6.4	58.0	25.0	C, B ₁ , B ₂ , B ₃ , B ₆ , B ₉	Ca, Fe, Mg, P, K, Na, Zn, Mn	[38,244,245]
Grass pea	-	24.4	2.8 (0.8)	55.94	11.4	C, B ₁ , B ₂	Ca, Fe, Mg, P, Zn	[155,221,246]
Guanacaste	-	33.9	2.8 (0.1)	56.8	1.3		Ca, Fe, Mg, K, Na, Zn, Cu	[236,247]
Horse gram	280	22	0.6	37.5	5.7		Ca, Fe, Mg, P, Zn, Mn, Cu	[248–250]
Housa groundnut	367	19–21	1.1	67–74	5.5	C, B ₁ , B ₂ , B ₃	Ca, Fe, P, K	[38,40]
Itching bean	382	27–37	6.6–8.8	46–53	6–10	C, B ₃	Ca, Fe, Mg, P, K, Na, Cu, Mn	[28,251,252]
Jack bean	389	21–27	3.5 (0.3)	60.6	2–8	-	Ca, Fe, Mg, P, K, Na, Zn, Cu, Mn, Pb	[40,253,254]
Kedaung		20.1	20 (13)		0.98		Ca, Fe, Mg, P, K, Na, Zn, Cu, Mn	[255,256]
Kidney bean	333	23.6	0.8 (0.1)	30.0	24.0	C, B ₁ , B ₂ , B ₃ , B ₉ , K, E	Ca, Fe, Mg, P, K	[38,237,257]
Lablab	344	21–29	1.7 (0.3)	60.74	25.6	B ₁ , B ₂ , B ₃ , B ₆ , B ₉	Ca, Fe, Mg, P, K, Na, Zn	[38,40,258,259]
Lima bean	338	21.5	0.7 (0.2)	63.4	19.0	B ₁ , B ₂ , B ₃ , B ₆ , B ₉ , K, E	Ca, Fe, Mg, P, K, Na, Zn	[38,230]
Morama bean	635	29–38	32–42 (10)	18.9	19–27	E	Ca, Fe, Mg, P, K, Na, Zn, Cu, Mn	[260,261]
Moth bean	343	23–26	1.6 (0.4)	61.5	5	C, B ₁ , B ₂ , B ₃ , B ₆ , B ₉ , A	Ca, Fe, Mg, P, K, Na, Zn	[38,40]
Mung bean	347	15–28	1.2 (0.4)	62.6	16.3	C, B ₁ , B ₂ , B ₃ , B ₆ , B ₉ , A	Ca, Fe, Mg, P, K, Na, Zn	[38,40,235,262,263]
Narbon bean	271	26.9	10–15	52–53			Ca, Fe, Mg, P, K, Zn, Cu, Mn, S	[241,264,265]
Navy bean	337	22.3	1.5 (0.2)	60.8	15.3	B ₁ , B ₂ , B ₃ , B ₆ , B ₉	Ca, Fe, Mg, P, K, Na, Zn	[38,266]
Pigeon pea	343	13–26	1.5 (0.3)	62.8	15	B ₁ , B ₂ , B ₃ , B ₆ , B ₉ , A	Ca, Fe, Mg, P, K, Na, Zn	[38,40]
Pinto bean	347	21.4	1.2 (0.2)	62.6	15.5	C, B ₉ , K, E	Ca, Fe, Mg, P, K, Na, Zn	[38,266]
Pinto peanut		27.1	49.7 (9.4)	21.4				[267]
Protein pea	352	23.8	1.2 (0.2)	63.7	25	C, B ₁ , B ₂ , B ₃ , B ₆ , B ₉ , A, E, K	Ca, Fe, Mg, P, K, Na, Zn	[38]
Purple mucuna	417	23.9	13.3 (5.3)	51.7	8.1	C, B ₃	Ca, Fe, Mg, P, K, Na, Zn, Cu, Mn	[268]
Red moneywort	439	16–27	14.0 (2.8)	54.6	4.25		Ca, Fe, Mg, P, K, Na, Zn, Cu, Mn	[269,270]
Rice bean	338	18–19	0.5 (0.3)	59.1	7.1	C, B ₂	Ca, Fe, P, Zn, Cu, Mn	[38,40,75,172]
Scarlet runner bean	338	20.3	1.8 (-)	62.0	4.8	C, B ₁ , B ₂ , B ₃	Ca, Fe, P	[231]
Sesban	459	30–40	5–6 (1–2)	45–47	11–16			[83,271]

Table A2. Cont.

Pulses—Common Name	Energy (kcal/100 g)	Protein (%)	Oil (Saturated FA) (%)	Carbohydrate (%)	Fiber (%)	Vitamins	Minerals	Reference
Sword bean	361	24–30	2.6–9.8 (-)	41–59	7–13	C, B ₁ , B ₂ ,	Ca, Fe, Mg, P, K, Na, Zn, Cu, Pb, Hg	[40,253,272,273]
Tamarind	239	24–25	8–12.5	10–19	3–4	C, B ₁ , B ₂ , B ₃	Ca, Fe, Mg, P, K, Na, Zn, Cu, Mn	[179,274–276]
Tarwi	440	41–51	14–24 (19)	28.2	7.1			[40,184,277]
Tepary bean	353	19–24	1.2 (-)	67.8	4.8	B ₁ , B ₂ , B ₃	Ca, Fe, P, K, Na	[40,231,262,278]
Urad	341	25.2	1.6 (0.1)	59.0	18.3	B ₁ , B ₂ , B ₃ , B ₆ , B ₉ , A	Ca, Fe, Mg, P, K, Na, Zn	[38]
Velvet bean	373	20–29	6–7	50–61	9–11		Ca, Fe, Mg, P, K, Na, Zn, Cu, Mn	[38,39,279–285]
Winged bean	428	30–35	16.3 (2.3)	41.7	11–26	B ₁ , B ₂ , B ₃ , B ₆ , B ₉	Ca, Fe, Mg, P, K, Na, Zn	[38,40,281]
Yam bean	390	10–32	24–26	31–33	7–8	C, B ₁ , B ₂ , B ₃ , B ₆	Ca, Fe, Mg, P, K, Na, Zn, Mn, Se	[72,286,287]
Broad bean	341	26.1	1.53 (0.3)	58.3	25	C, B ₁ , B ₂ , B ₃ , B ₆ , B ₉ , A, E, K	Ca, Fe, Mg, P, K, Na, Zn	[38,288]
Chickpea	378	11–31	6.0 (0.6)	63.0	12.2	C, B ₁ , B ₂ , B ₃ , B ₆ , B ₉ , A, E, K	Ca, Fe, Mg, P, K, Na, Zn	[38,40,289–291]
Common bean		20–24	0.8 (0.6)	75.5		B ₉ , A	Ca, Fe, Mg, K, P, Na, Zn, Mn, Se, S, B	[292–294]
Field pea	352	23.8	1.2 (0.2)	63.7	25	C, B ₁ , B ₂ , B ₃ , B ₆ , B ₉ , A, E, K	Ca, Fe, Mg, P, K, Na, Zn	[38]
Lentil	352	24.6	1.1 (0.2)	63.4	10.7	C, B ₁ , B ₂ , B ₃ , B ₆ , B ₉ , A, E, K	Ca, Fe, Mg, P, K, Na, Zn, Se	[38,295–297]
Lupin	371	36.2	9.7 (1.2)	40.37	18.9	C, B ₁ , B ₂ , B ₃ , B ₆ , B ₉	Ca, Fe, Mg, P, K, Na, Zn	[38,298]

Appendix C

Table A3. Nutritional composition of grains of non-pulses crops.

Other Crops—Common Name	Latin Name	Energy (Kcal)	Protein (%)	Oil (Saturated FA) (%)	Carbohydrate (%)	Fiber (%)	Vitamins	Minerals	References
African walnut	<i>Tetracarpidium conophorum</i>		30.1	43.4	16.9	2.6		Ca, Fe, Mg, K, Mn, Ni, Pb, Na, Cu	[299,300]
Almond	<i>Prunus dulcis</i>	579	21.2	49.9 (3.8)	21.6	12.5	B ₁ , B ₂ , B ₃ , B ₆ , B ₉ , E	Ca, Fe, Mg, P, K, Na, Zn	[38]
Cashew	<i>Anacardium occidentale</i>	552	18.2	43.9 (7.8)	30.2	3.3	C, B ₁ , B ₂ , B ₃ , B ₆ , B ₉ , E, K	Ca, Fe, Mg, P, K, Na, Zn	[38]
Castor bean	<i>Ricinus communis</i>	579	20.2	45.0		3.1		Ca, Fe, Mg, P, K, Na, Zn	[226]
Chia	<i>Salvia hispanica</i>	486	16.5	30.7 (3.3)	42.1	34.4	C, B ₁ , B ₂ , B ₃ , A, E	Ca, Fe, Mg, P, K, Na, Zn	[38]
Conophor nut	<i>Tetracarpidium conophorum</i>	590	22.8	49.02		5.5		Ca, Fe, Mg, P, K, Na, Zn	[226]
Cotton	<i>Gossypium hirsutum</i>		26–46	30–38		17.3		Ca, Mg, P, K, S	[301–303]
Cram Cram	<i>Cenchrus Biflorus</i>	370	17.8	8.5	62.3		B ₁ , B ₂ , B ₃	Ca, Fe, P	[231]
Egusi melon	<i>Citrullus colocynthis</i>	537	31.4	43.9		6.6	B ₁ , B ₂ , B ₃	Ca, Fe, Mg, P, K, Na, Zn, S	[226,304]
Flaxseed	<i>Linum usitatissimum</i>	376	24.4	30.9 (2.9)	0	38.6	B ₁ , B ₂ , B ₃ , K	Ca, Fe, Mg, P, K, Na, Zn, Mn	[305]

Table A3. Cont.

Other Crops—Common Name	Latin Name	Energy (Kcal)	Protein (%)	Oil (Saturated FA) (%)	Carbohydrate (%)	Fiber (%)	Vitamins	Minerals	References
Groundnut ^a	<i>Arachis hypogaea</i>	570	25.1	47.6	20.9	8.7	B ₁ , B ₂ , B ₃ , B ₆ , B ₉	Ca, Fe, Mg, P, K, Na, Zn	[38]
Hazelnut	<i>Corylus avellana</i>	628	15.0	60.8 (4.4)	19.2	11.2	C, B ₁ , B ₂ , B ₃ , B ₆ , B ₉ , A, E, K	Ca, Fe, Mg, P, K, Na, Zn	[38]
Hemp	<i>Cannabis sativa</i>	553	31.6	48.8 (4.6)	8.7	4.0	C, B ₁ , B ₂ , B ₃ , B ₆ , B ₉ , A, E	Ca, Fe, Mg, P, K, Na, Zn	[38]
Kalahari white bautinia	<i>Bautinia Petersiana</i>	371	22.9	13.1(3.0)	40.2	13.0	B ₁ , B ₂ , B ₃	Ca, Fe, P	[39]
Linseed	<i>Linum usitatissimum</i>	534	18.3	42.2 (3.6)	28.9	27.3	B ₁ , B ₂ , B ₃ , B ₅ , B ₆ , B ₉ , B ₁₂ , E, K	Ca, Fe, Ni, P, K, Na, Zn, Mn, Se, Co, Cu, Cr	[38]
Millet	<i>Pennisetum glaucum</i>	378	11.0	4.2 (0.7)	72.9	8.5	B ₁ , B ₂ , B ₃ , B ₆ , B ₉ , E, K	Ca, Fe, Mg, P, K, Na, Zn	[38]
Pistachio	<i>Pistacia Vera</i>	560	20.2	45.3 (5.9)	27.2	10.6	C, B ₁ , B ₂ , B ₃ , B ₆ , B ₉ , A, E	Ca, Fe, Mg, P, K, Na, Zn	[38]
Quinoa	<i>Chenopodium quinoa</i>	368	14.1	6.1 (0.7)	64.2	7.0	B ₁ , B ₂ , B ₃ , B ₆ , B ₉ , A, E	Ca, Fe, Mg, P, K, Na, Zn	[38]
Rapeseed	<i>Brassica napus</i>		18–20	43–45		23–27			[306,307]
Sesame	<i>Sesamum indicum</i>	573	17.7	49.7 (7.0)	23.5	11.8	B ₁ , B ₂ , B ₃ , B ₆ , B ₉ , A, E	Ca, Fe, Mg, P, K, Na, Zn	[38]
Soybean	<i>Glycine Mac Merrill</i>	446	36.5	19.9 (2.9)	30.2	9.3	C, B ₁ , B ₂ , B ₃ , B ₆ , B ₉ , A, E, K	Ca, Fe, Mg, P, K, Na, Zn	[38]
Spirulina (dried)	<i>Arthrospira maxima</i>	290	57.5	7.7 (2.7)	23.9	3.6	C, B ₁ , B ₂ , B ₃ , B ₆ , B ₉ , A, E, K	Ca, Fe, Mg, P, K, Na, Zn	[38]
Sunflower (kernel)	<i>Helianthus annuus</i>	584	18–21	51.0 (4.4)	20	8.6	C, B ₁ , B ₂ , B ₃ , B ₆ , B ₉ , A, E	Ca, Fe, Mg, P, K, Na, Zn, Se	[38]
Walnut	<i>Juglans spp.</i>	654	15.2	65.2 (6.1)	13.7	6.7	C, B ₁ , B ₂ , B ₃ , B ₆ , B ₉ , A, E, K	Ca, Fe, Mg, P, K, Na, Zn	[38]

^a Equivalent to peanut.

Appendix D

Table A4. Antinutritional constituents of pulses and other crops with high protein content, in grey major pulses.

Crops—Common Name	Phytic Acid (mg/g)	Tannins Total (mg/g)	Saponins (0)	Trypsin Inhibitors (mg/100 g)	Lectins (mg/100 g)	Others ^a	References
Acacia leucophloea	Presence	0.01		Presence			[221]
Adzuki bean		2.9					[308]
African locust bean	0.6	0.81		19.4		HCN; OX	[224–226]

Table A4. Cont.

Crops— Common Name	Phytic Acid (mg/g)	Tannins Total (mg/g)	Saponins (μ)	Trypsin Inhibitors (mg/100 g)	Lectins (mg/100 g)	Others ^a	References
African nut bean	Presence	0.07–0.3				OX	[309]
African oil bean	41	7.9	17.8			HCN	[99,226]
African yam bean	4.3–14.9	18.1	1.2	6.7		HCN; OX	[79,310]
Bambara groundnut	0.5–14.8	tr-5.0	1.4	6.7		OX	[232,311–313]
Butterfly pea	11.5	8.7				HCN; L-DOPA ⁱ	[236]
Castor bean	0.89	0.11				OX; A	[226]
Conophor nut	2.1	0.21				OX	[226]
Cotton		0.1				GO	[302]
Cowpea	1.4–3.8	1.4–10.2	0.3	26.4 ^b		HCN; OX	[82,311,314, 315]
Egusi melon	4.1	0.8				OX	[226]
Ervil		Presence		Presence		PA	[239,241]
Fenugreek			0.1–0.9				[244,245]
Grass pea	3.0	0.2–0.8		19.64 ^b		β -ODAP; CTI	[246,316–319]
Groundnut ^h	4.18	0.04		80.8 ^b		OX; HCN; AL; AI	[87]
Guanacaste	9.5	3.7				HCN; L-DOPA	[236]
Horse gram		2		865			[248,250]
Housa groundnut	Presence	Presence			Presence		[320]
Itching bean	4.7–56.8	1.8–3.3	1.2–1.3	43.2–43.7 ^b		HCN	[28,251,311]
Jack bean	12.0–13.7	Tr-0.7	1.8	16.4 ^b		HCN; CV; OX	[230,311,314, 321,322]
Kedaung		98.3	0.3	6.94 ^b			[81,142]
Kidney bean	17.3–24.1	5.4–28.8	0.9–23	4.6–29.3 ^b	1.92–9.98 ^c	HCN; OX	[82,323]
Lablab	6.1–15.7	0.2–0.4	1.3	19.7 ^b		OX	[230,258,311, 314,322]
Lima bean	13.6	6.5–9.1	1.2–1.5	2.1–17.2 ^b		HCN; OX	[230,308,311, 322]
Linseed						HCN	[324]
Morama bean			0.08	Presence			[263,325]
Moth bean	3.8–4.2	4.8–13	33	28.3–31.4 ^b		HCN	[82,311]
Mung bean	5.8–7.4	4.4–8.0	2.8–35	15.8 ^b	2670 ^d	OX	[82,84,308, 326]
Narbon bean		Presence		Presence		PA	[239,241,327]

Table A4. Cont.

Crops—Common Name	Phytic Acid (mg/g)	Tannins Total (mg/g)	Saponins (0)	Trypsin Inhibitors (mg/100 g)	Lectins (mg/100 g)	Others ^a	References
Navy bean	12.9–15.8	39.9	20–160	5.9 ^f	3.8 ^c	OX	[82,328–330]
Pigeon pea	7.3–16.2	3.8–17.1	0.04–1.4	4.1–19.2 ^b		HCN; OX; CTI; PA	[40,82,230,311,331]
Pinto bean		2.6			2.3 ^c	OX	[82,266,308,330]
Pinto peanut		Presence					
Purple mucuna	3.8–4.5	1.8–3.4		39.2–44.1		HCN	[311]
Rapeseed	Presence	Presence				Glucosinolates	[324]
Red moneywort		Presence					[269]
Rice bean	3.3–20.3	2.4	2.3	34.3–40.6	Tr	HCN	[75,172,311,332]
Scarlet runner bean				Presence [*]	Presence [*]		[333,334]
Sesban	18–51	19	5.2–14.6	50–140	Presence	PA	[83,272]
Sword bean	3.5–21.4	0.01–570	1.7–5.2	17.4–26.8 ^b		HCN; OX	[80,230,311,322,335]
Tamarind	Presence	Presence		Presence			[180,336]
Tarwi				T		AL; HCN	[184]
Tepary bean			1–37	11.5–18.0 ^b	1.4–18.2 ^e		[82]
Urad	11.2–14.6	5.4–11.9	0.2–23	94.2 ^g			[82,308,314,337]
Velvet bean	5.0–53.6	1.8–3.1		43.2–52.8		AL; HCN; L-DOPA	[28,311,321,338–342]
Winged bean	7.8–12.3	0.3–12.6		0.01–0.14 ^e	76 ^e	CTI	[343,344]
Yam bean	Tr	Tr		0.01	0.0003	HCN	[287]
Broad bean	6.4	0.1–24.1	0.4–370	1.7–3.3 ^b	25–100 ^d	CTI; L-DOPA	[82,87,311,345]
Chickpea	1.2–12.1	08–5.9	0.09–600	11.9 ^b	6.22 ^d	HCN; OX; PA; CTI	[40,82,311,346]
Common bean	8.2			0–4.64 ^g	8.57	CTI; AI	[82,294,347]
Field pea	2.2–7.4	0.2–13	18–110	1.5–108	5.1–150.6	OS; PA; AI; HCN; CTI	[82,348]
Lentil	2.4–12.4	12.8	0.04–0.13	2.8		OX; AI	[349,350]
Lupin	1.4–3.5	ABS	ABS	ABS	ABS	PH; AL	[351–353]
Soybean	10.0–23.0		0.2–5.6	0.2–1.12 ^b			[230,311,354]

^a Presence of: OS: oligosaccharides; PA: phenolic acid; AI: amylase inhibitor; HCN: hydrogen cyanide; CV: canavanine; OX: oxalate; A: allergens; PH: phomopsine; GO: gossypol; CTI: chymotrypsin inhibitors; GC: γ -glutamyl- β -cyanialanine; VC3: vicianine + vicine + convicine; CA: caravanine; AL: alkaloids; β -ODAP (beta-oxalyl-diamino-propionic acid); TAN: tannins; ^b: units trypsin units inhibited TUI/mg protein; ^c: units g/kg PHA: lectin as PHA (*P. vulgaris* lectin); ^d: units HU/mg sample; ^e: units HUA/g proteins; ^f: units TIA/g sample; ^g: units IU/g seed; ^h: data reported in groundnut oil; ⁱ 1-3,4-dihydroxyphenylalanin; ND: non-detected; Tr: traces; ABS: absence; * presence: molecules detected but unquantified.

References

- De Boer, J.; Aiking, H. On the merits of plant-based proteins for global food security: Marrying macro and micro perspectives. *Ecol. Econ.* **2011**, *70*, 1259–1265. [CrossRef]
- Ranalli, P. Improvement of pulse crops in Europe. *Eur. J. Agron.* **1995**, *4*, 151–166. [CrossRef]
- Voisin, A.-S.; Guéguen, J.; Huyghe, C.; Jeuffroy, M.-H.; Magrini, M.-B.; Meynard, J.-M.; Mougél, C.; Pellerin, S.; Pelzer, E. Legumes for feed, food, biomaterials and bioenergy in Europe: A review. *Agron. Sustain. Dev.* **2013**, *34*, 361–380. [CrossRef]
- Smykal, P.; Coyne, C.J.; Ambrose, M.J.; Maxted, N.; Schaefer, H.; Blair, M.W.; Berger, J.; Greene, S.L.; Nelson, M.N.; Besharat, N.; et al. Legume crops phylogeny and genetic diversity for science and breeding. *Crit. Rev. Plant Sci.* **2015**, *34*, 43–104. [CrossRef]
- Ayerdi Gotor, A.; Marraccini, E. Crops providing proteins for food: A review. In Proceedings of the 14th ESA Congress—“Growing Landscapes—Cultivating Innovative Agricultural Systems”, Edinburgh, UK, 5–9 September 2016; pp. 27–28.
- FAOSTAT. Available online: <https://www.fao.org/faostat/en/#data> (accessed on 10 January 2021).
- Ahlawat, I.; Sharma, P.; Singh, U. Production, demand and import of pulses in India. *Indian J. Agron.* **2016**, *4*, S33–S41.
- Watson, C.A.; Reckling, M.; Preissel, S.; Bachinger, J.; Bergkvist, G.; Kuhlman, T.; Lindström, K.; Nemecek, T.; Topp, C.F.E.; Vanhatalo, A.; et al. Chapter Four—Grain legume production and use in European agricultural systems. In *Advances in Agronomy*; Sparks, D.L., Ed.; Academic Press: Cambridge, MA, USA, 2017; Volume 144, pp. 235–303. ISBN 0065-2113.
- Kearney, J. Food consumption trends and drivers. *Philos. Trans. R. Soc. B Biol. Sci.* **2010**, *365*, 2793–2807. [CrossRef]
- Schneider, A.V.C. Overview of the market and consumption of pulses in Europe. *Br. J. Nutr.* **2002**, *88*, 243–250. [CrossRef]
- Desriers, M. L’agriculture française depuis cinquante ans: Des petites exploitations familiales aux droits à paiement unique. *Agreste Cah.* **2007**, *2*, 3–14.
- European Commission Development of Plant Proteins in the European Union. *Report*; Commission to the Council/The European Parliament: Brussels, Belgium, 2018; p. 16.
- Wezel, A.; Soboksa, G.; McClelland, S.; Delespesse, F.; Boissau, A. The blurred boundaries of ecological, sustainable, and agroecological intensification: A review. *Agron. Sustain. Dev.* **2015**, *35*, 1283–1295. [CrossRef]
- Bowen, C.R.; Hollinger, S.E. Geographic screening of potential alternative crops. *Renew. Agric. Food Syst.* **2004**, *19*, 141–151. [CrossRef]
- Von Richthofen, J.-S.; Pahl, H.; Bouttet, D.; Casta, P.; Cartryse, C.; Charles, R.; Lafarga, A. What do European farmers think about grain legumes? *Grain Legumes* **2006**, *45*, 14–15.
- Zander, P.; Amjath-Babu, T.S.; Preissel, S.; Reckling, M.; Bues, A.; Schläfke, N.; Kuhlman, T.; Bachinger, J.; Uthes, S.; Stoddard, F.; et al. Grain legume decline and potential recovery in European agriculture: A review. *Agron. Sustain. Dev.* **2016**, *36*, 26. [CrossRef]
- Magrini, M.-B.; Anton, M.; Cholez, C.; Corre-Hellou, G.; Duc, G.; Jeuffroy, M.-H.; Meynard, J.-M.; Pelzer, E.; Voisin, A.-S.; Walrand, S. Why are grain-legumes rarely present in cropping systems despite their environmental and nutritional benefits? Analyzing lock-in in the French agrifood system. *Ecol. Econ.* **2016**, *126*, 152–162. [CrossRef]
- Reckling, M.; Döring, T.; Bergkvist, G.; Stoddard, F.; Watson, C.; Seddig, S.; Chmielewski, F.-M.; Bachinger, J. Grain legume yields are as stable as other spring crops in long-term experiments across northern Europe. *Agron. Sustain. Dev.* **2018**, *38*, 1–10. [CrossRef] [PubMed]
- Chimonyo, V.G.P.; Snapp, S.; Chikowo, R. Grain legumes increase yield stability in maize-based cropping systems. *Crop Sci.* **2019**, *59*, 1222–1235. [CrossRef]
- Mawois, M.; Vidal, A.; Revoyron, E.; Casagrande, M.; Jeuffroy, M.-H.; Le Bail, M. Transition to legume-based farming systems requires stable outlets, learning, and peer-networking. *Agron. Sustain. Dev.* **2019**, *39*, 1–14. [CrossRef]
- Chloupek, O.; Hrstkova, P. Adaptation of crops to environment. *Theor. Appl. Genet.* **2005**, *111*, 1316–1321. [CrossRef]
- Udendi, O.; Umana, E.; Edu, E.; Ikpeme, E. Screening locally grown pulses for proximate, anti-nutritive and mineral compositions: Indices for conservation and improvement. *Int. J. Agric. Res.* **2011**, *6*, 504–510. [CrossRef]
- Jacobsen, S.-E. The worldwide potential for quinoa (*Chenopodium quinoa* Willd.). *Food Rev. Int.* **2003**, *19*, 167–177. [CrossRef]
- Bazile, D.; Jacobsen, S.-E.; Verniau, A. The global expansion of quinoa: Trends and limits. *Front. Plant Sci.* **2016**, *7*, 622. [CrossRef]
- Więcicki, W.; Świącicki, W.K.; Wolko, B. *Lupinus anatolicus*—A new lupin species of the old world. *Genet. Resour. Crop Evol.* **1996**, *43*, 109–117.
- Spataro, G.; Tiranti, B.; Arcaleni, P.; Bellucci, E.; Attene, G.; Papa, R.; Zeuli, P.S.; Negri, V. Genetic diversity and structure of a worldwide collection of *Phaseolus coccineus* L. *Theor. Appl. Genet.* **2011**, *122*, 1281–1291. [CrossRef] [PubMed]
- Srivastava, K.M.; Singh, L.N. A review of the pest complex of Kharif pulses in Uttar Pradesh. *PANS* **1976**, *22*, 333–335. [CrossRef]
- Tresina, P.S.; Mohan, V.R. Assessment of nutritional and antinutritional potential of underutilized legumes of the Genus *mucuna*. *Trop. Subtrop. Agroecosyst.* **2013**, *16*, 155–169.
- Rana, J.C.; Gautam, N.K.; Gayacharan, M.S.; Yadav, R.; Tripathi, K.; Yadav, S.K.; Panwar, N.S.; Bhardwaj, R. Genetic resources of pulse crops in India: An overview. *Indian J. Genet. Plant Breed.* **2016**, *76*, 420. [CrossRef]
- Thomson, B.; Siddique, K.; Barr, M.; Wilson, J. Grain legume species in low rainfall Mediterranean-type environments I. Phenology and seed yield. *Field Crop. Res.* **1997**, *54*, 173–187. [CrossRef]
- Saka, J.O.; Ajibade, S.R.; Adeniyani, O.N.; Olowoyo, R.B.; Ogunbodede, B.A. Survey of underutilized grain legume production systems in the southwest agricultural zone of Nigeria. *J. Agric. Food Inf.* **2004**, *6*, 93–108. [CrossRef]

32. Azam-Ali, S. Agricultural diversification: The potential for underutilised crops in Africa's changing climates. *Riv. Biol. Biol. Forum* **2007**, *1*, 27–28. [CrossRef]
33. Kaoneka, S.R.; Saxena, R.K.; Silim, S.N.; Odeny, D.A.; Rao, N.V.; Shimelis, H.A.; Siambi, M.; Varshney, R.K. Pigeonpea breeding in eastern and southern Africa: Challenges and opportunities. *Plant Breed.* **2016**, *135*, 148–154. [CrossRef]
34. Vadivel, V.; Janardhanan, K. Nutritional and antinutritional characteristics of seven south Indian wild legumes. *Plant Foods Hum. Nutr.* **2005**, *60*, 69–75. [CrossRef]
35. Vadivel, V.; Biesalski, H.K. Bioactive compounds in velvet bean seeds: Effect of certain indigenous processing methods. *Int. J. Food Prop.* **2012**, *15*, 1069–1085. [CrossRef]
36. Pullin, A.S.; Stewart, G.B. Guidelines for systematic review in conservation and environmental management. *Conserv. Biol.* **2006**, *20*, 1647–1656. [CrossRef] [PubMed]
37. Elsevier. *Scopus® Quick Reference Guide*; Elsevier: Amsterdam, The Netherlands, 2019; p. 12.
38. USDA. Food Composition Database. 2018. Available online: <https://ndb.nal.usda.gov/ndb/> (accessed on 28 October 2018).
39. PROTA. *Céréales et Légumes Secs*; PROTA: Ankara, Turkey, 2006; ISBN 978-90-5782-172-1.
40. FAO. *Utilisation Des Aliments Tropicaux: Légumineuses Tropicales (47/4)*; *Cahiers Techniques de la FAO*; Food and Agriculture Organization of the United Nations: Rome, Italy, 1990.
41. Page, M.J.; McKenzie, J.E.; Bossuyt, P.M.; Boutron, I.; Hoffmann, T.C.; Mulrow, C.D.; Shamseer, L.; Tetzlaff, J.M.; Akl, E.A.; Brennan, S.E.; et al. The PRISMA 2020 statement: An updated guideline for reporting systematic reviews. *BMJ* **2021**, *372*, n71. [CrossRef] [PubMed]
42. Giraldeli, A.; Gregorio, J.S.; Monquero, P.; Aguilera, M.; Ribeiro, N. Weeds hosts of nematodes in sugarcane culture. *Planta Daninha* **2017**, *35*. [CrossRef]
43. Correia, M.V.; Pereira, L.C.; De Almeida, L.; Williams, R.L.; Freach, J.; Nesbitt, H.; Erskine, W. Maize mucuna (*Mucuna pruriens* (L.) DC) relay intercropping in the lowland tropics of Timor-Leste. *Field Crop. Res.* **2014**, *156*, 272–280. [CrossRef]
44. Mohamed, H.I.; Hameed, A.G.A.-E. Molecular and biochemical markers of some *Vicia faba* L. genotypes in response to storage insect pests infestation. *J. Plant Interact.* **2014**, *9*, 618–626. [CrossRef]
45. Oliveira, H.R.; Tomás, D.; Silva, M.; Lopes, S.; Viegas, W.; Veloso, M.M. Genetic diversity and population structure in *Vicia faba* L. landraces and wild related species assessed by nuclear SSRs. *PLoS ONE* **2016**, *11*, e0154801. [CrossRef]
46. Hanly, J.A.; Gregg, P.E.H. Green-manure impacts on nitrogen availability to organic sweetcorn (*Zea mays*). *N. Z. J. Crop. Hortic. Sci.* **2004**, *32*, 295–307. [CrossRef]
47. Jayanthi, M.; Umarani, R.; Vijayalakshmi, V. Effect of seed fortification with pulse sprout extract on crop growth and seed yield in rice seeds. *Asian J. Crop. Sci.* **2013**, *5*, 444–451. [CrossRef]
48. Caracuta, V.; Vardi, J.; Paz, Y.; Boaretto, E. Farming legumes in the pre-pottery Neolithic: New discoveries from the site of Ahihud (Israel). *PLoS ONE* **2017**, *12*, e0177859. [CrossRef] [PubMed]
49. Lykke, A.M.; Mertz, O.; Ganaba, S. Food consumption in rural Burkina Faso. *Ecol. Food Nutr.* **2002**, *41*, 119–153. [CrossRef]
50. Malau-Aduli, B.S.; Eduvie, L.; Lakpini, C.; Malau-Aduli, A.E.O. Crop-residue supplementation of pregnant does influences birth weight and weight gain of kids, daily milk yield but not the progesterone profile of Red Sokoto goats. *Reprod. Nutr. Dev.* **2004**, *44*, 111–121. [CrossRef]
51. Pastor-Cavada, E.; Juan, R.; Pastor, J.E.; Alaiz, M.; Girón-Calle, J.; Vioque, J. Antioxidative activity in the seeds of 28 *Vicia* species from southern Spain. *J. Food Biochem.* **2011**, *35*, 1373–1380. [CrossRef]
52. Cernay, C.; Pelzer, E.; Makowski, D. A global experimental dataset for assessing grain legume production. *Sci. Data* **2016**, *3*, 160084. [CrossRef]
53. Tancoigne, E.; Barbier, M.; Cointet, J.-P.; Richard, G. The place of agricultural sciences in the literature on ecosystem services. *Ecosyst. Serv.* **2014**, *10*, 35–48. [CrossRef]
54. Ruiz-Martinez, I.; Marraccini, E.; Debolini, M.; Bonari, E. Indicators of agricultural intensity and intensification: A review of the literature. *Ital. J. Agron.* **2015**, *10*, 74. [CrossRef]
55. Metzger, M.J.; Bunce, R.G.H.; Jongman, R.H.G.; Múcher, C.A.; Watkins, J.W. A climatic stratification of the environment of Europe. *Glob. Ecol. Biogeogr.* **2005**, *14*, 549–563. [CrossRef]
56. Major, D.J.; Brown, D.M.; Bootsma, A.; Dupuis, G.; Fairey, N.A.; Grant, E.A.; Green, D.G.; Hamilton, R.I.; Langille, J.; Sonmor, L.G.; et al. An evaluation of the corn heat unit system for the short season growing regions across Canada. *Can. J. Plant Sci.* **1983**, *63*, 121–130. [CrossRef]
57. Ray, H.; Bett, K.; Tar'An, B.; Vandenberg, A.; Thavarajah, D.; Warkentin, T. Mineral micronutrient content of cultivars of field pea, chickpea, common bean, and lentil grown in Saskatchewan, Canada. *Crop Sci.* **2014**, *54*, 1698–1708. [CrossRef]
58. Ribeiro, H.L.C.; Boiteux, L.S.; Santos, C.A.F. Genetic parameters of earliness and plant architecture traits suitable for mechanical harvesting of cowpea (*Vigna unguiculata*). *Aust. J. Crop Sci.* **2014**, *8*, 1232–1238.
59. De Oliveira, J.T.; Ribeiro, I.D.S.; Roque, C.G.; Montanari, R.; Gava, R.; Teodoro, P.E. Contribution of morphological traits for grain yield in common bean. *Biosci. J.* **2018**, *34*, 951–956. [CrossRef]
60. Alliprandini, L.F.; Abatti, C.; Bertagnolli, P.F.; Cavassim, J.E.; Gabe, H.L.; Kurek, A.; Matsumoto, M.N.; De Oliveira, M.A.R.; Pitol, C.; Prado, L.C.; et al. Understanding soybean maturity groups in Brazil: Environment, cultivar classification, and stability. *Crop. Sci.* **2009**, *49*, 801–808. [CrossRef]

61. Jia, H.; Jiang, B.; Wu, C.; Lu, W.; Hou, W.; Sun, S.; Yan, H.; Han, T. Maturity group classification and maturity locus genotyping of early-maturing soybean varieties from high-latitude cold regions. *PLoS ONE* **2014**, *9*, e94139. [[CrossRef](#)] [[PubMed](#)]
62. Appunu, C.; N'Zoue, A.; Moulin, L.; Depret, G.; Laguerre, G. *Vigna mungo*, *V. radiata* and *V. unguiculata* plants sampled in different agronomical–ecological–climatic regions of India are nodulated by *Bradyrhizobium yuanmingense*. *Syst. Appl. Microbiol.* **2009**, *32*, 460–470. [[CrossRef](#)] [[PubMed](#)]
63. Bouget, M. *Les Légumineuses Vivrières. Le Technicien d'agriculture Tropicale*; Editions Maisonneuve et Larose: Paris, France, 1989; 162p.
64. Sørensen, M.; Heller, J.; Engels, J. *Yam Bean. Pachyrhizus DC.—Promoting the Conservation and Use of Underutilized and Neglected Crops*; International Plant Genetic Resources Institute: Rome, Italy, 1996; Volume 2, ISBN 978-92-9043-282-1.
65. Klimek-Kopyra, A.; Kulig, B.; Oleksy, A.; Zajac, T. Agronomic performance of naked oat (*Avena nuda* L.) and faba bean intercropping. *Chil. J. Agric. Res.* **2015**, *75*, 168–173. [[CrossRef](#)]
66. Akobundu, I.O.; Udensi, U.E.; Chikoye, D. Velvetbean (*Mucuna* spp.) suppresses speargrass (*Imperata cylindrica* (L.) Raeuschel) and increases maize yield. *Int. J. Pest Manag.* **2000**, *46*, 103–108. [[CrossRef](#)]
67. Paulo, E.M.; Berton, R.S.; Cavichioli, J.C.; Bulisani, E.A.; Kasai, F.S. Produtividade do cafeeiro Mundo Novo enxertado e submetido à adubação verde antes e após recepa da lavoura. *Bragantia* **2006**, *65*, 115–120. [[CrossRef](#)]
68. Lusembo, P.; Ebong, C.; Sabiiti, E.N. Integration of cassava tuber and forage legume seed production for sustained soil fertility. *Trop. Agric.* **1998**, *75*, 18–20.
69. Ribeiro, G.M.; Neto, F.B.; De Lima, J.S.S.; Da Silva, M.L.; Júnior, A.P.B.; Dos Santos, E.C. Productive performance of carrot and cowpea intercropping system under different spatial arrangements and population densities. *Rev. Caatinga* **2018**, *31*, 19–27. [[CrossRef](#)]
70. Ocimati, W.; Ntamwira, J.; Groot, J.; Taulya, G.; Tittonell, P.; Dhed'A, B.; van Asten, P.; Vanlauwe, B.; Ruhigwa, B.; Blomme, G. Banana leaf pruning to facilitate annual legume intercropping as an intensification strategy in the East African highlands. *Eur. J. Agron.* **2019**, *110*, 125923. [[CrossRef](#)]
71. Ameh, G. Proximate and mineral composition of seed and tuber of African yam bean, *Sphenostylis stenocarpa* (Hoechst. Ex. A. Rich.) harms. *ASSET Ser. B* **2007**, *6*, 1–10.
72. Kisambira, A.; Muyonga, J.H.; Byaruhanga, Y.B.; Tukamuhabwa, P.; Tumwegamire, S.; Grüneberg, W.J. Composition and functional properties of yam bean (*Pachyrhizus* spp.) seed flour. *Food Nutr. Sci.* **2015**, *06*, 736–746. [[CrossRef](#)]
73. Hardarson, G. Methods for enhancing symbiotic nitrogen fixation. *Plant Soil* **1993**, *152*, 1–17. [[CrossRef](#)]
74. Liu, J.; You, L.; Amini, M.; Obersteiner, M.; Herrero, M.; Zehnder, A.J.B.; Yang, H. A high-resolution assessment on global nitrogen flows in cropland. *Proc. Natl. Acad. Sci. USA* **2010**, *107*, 8035–8040. [[CrossRef](#)]
75. Bepary, R.H.; Wadikar, D.D.; Patki, P.E. Rice bean: Nutritional vibrant bean of Himalayan belt (Northeast India). *Nutr. Food Sci.* **2016**, *46*, 412–431. [[CrossRef](#)]
76. Spencer, P.F.; Schaumburg, H.H. Lathyrism: A neurotoxic disease. *Neurobehav. Toxicol. Teratol.* **1983**, *5*, 625–629. [[PubMed](#)]
77. Franco, F. *Decreto 2484/1967, de 21 de Septiembre*; El Código Alimentario Español (CAE): Barcelona, Spain, 1967; Volume 8.
78. Campbell, C.G.; Mehra, R.B.; Agrawal, S.K.; Chen, Y.Z.; El Moneim, A.M.A.; Khawaja, H.I.T.; Yadov, C.R.; Tay, J.U.; Araya, W.A. Current status and future strategy in breeding grasspea (*Lathyrus sativus*). *Euphytica* **1993**, *73*, 167–175. [[CrossRef](#)]
79. Ndidi, U.S.; Ndidi, C.U.; Olagunju, A.; Muhammad, A.; Billy, F.G.; Okpe, O. Proximate, antinutrients and mineral composition of raw and processed (boiled and roasted) *Sphenostylis stenocarpa* seeds from southern Kaduna, northwest Nigeria. *ISRN Nutr.* **2014**, *2014*, 1–9. [[CrossRef](#)] [[PubMed](#)]
80. Abitogun, A.; Oso, G.K. Assessment of processing methods on the chemical composition of sword bean (*Canavaliagradiata*). *IOSR J. Appl. Chem.* **2014**, *7*, 106–112. [[CrossRef](#)]
81. Sala, J.S.; Salam, P.; Pots, K.S.; Kuma, D.B. Effect of processing methods on secondary metabolites and enzyme inhibitors in different developmental stages of *Parkia roxburghii* G. Don Pods. *Am. J. Food Technol.* **2014**, *9*, 89–96. [[CrossRef](#)]
82. Campos-Vega, R.; Loarca-Piña, G.; Oomah, B.D. Minor components of pulses and their potential impact on human health. *Food Res. Int.* **2010**, *43*, 461–482. [[CrossRef](#)]
83. Hossain, M.; Becker, K. Nutritive value and antinutritional factors in different varieties of Sesbania seeds and their morphological fractions. *Food Chem.* **2001**, *73*, 421–431. [[CrossRef](#)]
84. Kataria, A.; Chauhan, B.; Punia, D. Antinutrients and protein digestibility (in vitro) of mungbean as affected by domestic processing and cooking. *Food Chem.* **1989**, *32*, 9–17. [[CrossRef](#)]
85. Inuwa, H.M.; Aina, V.O.; Gabi, B.; Aimola, I.; Toyin, A. Comparative determination of antinutritional factors in groundnut oil and palm oil. *Adv. J. Food Sci. Technol.* **2011**, *5*, 275–279.
86. Phillipson, J.D. *Natural Toxicants in Feeds and Poisonous Plants*; Cheeke, P.R., Shull, L.R., Eds.; AVI Publishing Company: Westport, CT, USA, 1985; 492p, ISBN 0870554824.
87. El-Hady, E.A.; Habiba, R. Effect of soaking and extrusion conditions on antinutrients and protein digestibility of legume seeds. *LWT* **2003**, *36*, 285–293. [[CrossRef](#)]
88. Agroforestry Database 4.0. *World Agroforestry Acacia leucophloea*; World Agroforestry: Nairobi, Kenya, 2019; p. 5.
89. Lumpkin, T.A.; McClary, D.C. *Azuki Bean: Botany, Production and Uses*; CAB International: Wallingford, UK, 1994; ISBN 0-85198-765-6.
90. Haq, N. Underutilized field legumes. In *Biology and Breeding of Food Legumes*; Pratap, A., Kumar, J., Eds.; CABI: Oxfordshire, UK, 2011; pp. 329–347.

91. Han, L.L.; Wang, E.T.; Lu, Y.L.; Zhang, Y.F.; Sui, X.H.; Chen, W.F.; Chen, W.X. *Bradyrhizobium* spp. and *Sinorhizobium fredii* are predominant in root nodules of *Vigna angularis*, a native legume crop in the subtropical region of China. *J. Microbiol.* **2009**, *47*, 287–296. [[CrossRef](#)] [[PubMed](#)]
92. Singh, R.J.; Jauhar, P.P. Grain legumes. In *Genetic Resources, Chromosome Engineering, and Crop Improvement*; Taylor & Francis: Boca Raton, FL, USA, 2005; Volume 1, ISBN 0849314305.
93. Fujihara, S.; Terakado, J.; Takenaka, M.; Yoneyama, T. Specific occurrence of β -phenethylamine in root nodules formed from *Bradyrhizobium*-legume symbiosis. *Plant Soil* **2002**, *238*, 123–132. [[CrossRef](#)]
94. Kang, Y.J.; Lee, J.; Kim, Y.H.; Lee, S.-H. Identification of tissue-specific gene clusters and orthologues of nodulation-related genes in *Vigna angularis*. *Plant Genet. Resour.* **2014**, *12*, S21–S26. [[CrossRef](#)]
95. Kumar, R.; Mittal, R.K.; Pandey, D.P. Genetic variability for yield and growth attributes in adzuki bean. *Res. Crops* **2012**, *13*, 562–565.
96. Wang, S.M.; Redden, R.J.; Jiapeng, J.P.H.; Desborough, P.J.; Lawrence, P.L.; Usher, T. Chinese adzuki bean germplasm: 1. Evaluation of agronomic traits. *Aust. J. Agric. Res.* **2001**, *52*, 671–681. [[CrossRef](#)]
97. Van Oers, C.C.C.M. *Vigna angularis* (Willd.) Ohwi & Ohashi. In *Pulses; Plant Resources of South-East, Asia*; van der Maesen, L.J.G., Somaatmadja, S., Eds.; PROTA: Bogor, Indonesia, 1989.
98. Feedipedia. List of Feeds. 2020. Available online: <https://www.feedipedia.org/content/feeds?category=13596> (accessed on 26 November 2018).
99. Janick, J.; Paull, R.E. The Encyclopedia of Fruit & Nuts. 2008. Available online: <http://www.credoreference.com/book/cabifruit> (accessed on 5 May 2020).
100. Lewin, A.; Rosenberg, C.; Meyer, Z.A.H.; Wong, C.H.; Nelson, L.; Manen, J.-F.; Stanley, J.; Dowling, D.; Denarie, J.; Broughton, W.J. Multiple host-specificity loci of the broad host-range *Rhizobium* sp. NGR234 selected using the widely compatible legume *Vigna unguiculata*. *Plant Mol. Biol.* **1987**, *8*, 447–459. [[CrossRef](#)] [[PubMed](#)]
101. Heller, J.; Begemann, F.; Mushonga, J. *Bambara Groundnut Vigna subterranea* (L.) Verdc: *Proceedings of the Workshop on Conservation and Improvement of Bambara Groundnut (Vigna subterranea* (L.) Verdc.): *Promoting the Conservation and Use of Underutilized and Neglected Crops*; International Plant Genetic Resources Institute: Harare, Zimbabwe, 1997; ISBN 978-92-9043-299-9.
102. Verheye, W.H. *Soils, Plant Growth and Crop Production*; EOLSS Publications: Abu Dhabi, United Arab Emirates, 2010; Volume 3, ISBN 978-1-84826-369-7.
103. Alake, C.O.; Ayo-Vaughan, M.A.; Ariyo, J.O. Selection criteria for grain yield and stability in Bambara groundnut (*Vigna subterranean* (L) Verdc) landraces. *Acta Agric. Scand. Sect. B Soil Plant Sci.* **2015**, *65*, 433–447. [[CrossRef](#)]
104. Alake, C.O. Genetic variability, gains from selection and genetic correlations for pod yield and nutritional traits in African landraces of Bambara groundnut. *Biol. Agric. Horticult.* **2017**, *34*, 71–87. [[CrossRef](#)]
105. Berchie, J.; Sarkodie-A, J.; Adu-Dapaah, H.; Agyemang, A.; Addy, S.; Asare, E.; Donkor, J. Yield evaluation of three early maturing Bambara groundnut (*Vigna subterranea* L. Verdc) landraces at the CSIR—Crops Research Institute, Fumesua-Kumasi, Ghana. *J. Agron.* **2010**, *9*, 175–179. [[CrossRef](#)]
106. Hasan, M.; Uddin, K.; Mohamed, M.T.M.; Zuan, A.T.K. Nitrogen and phosphorus management for Bambara groundnut (*Vigna subterranea*) production—A review. *Legum. Res. Int. J.* **2018**, *41*, 483–489. [[CrossRef](#)]
107. Wassermann, V.D.; Kruger, A.J.; Heyns, G. The response of Bambara groundnut (*Voandzeia subterranea*) and pigeon pea (*Cajanus cajan*) to applications of lime, P and K. *S. Afr. J. Plant Soil* **1984**, *1*, 4–8. [[CrossRef](#)]
108. Thottappilly, G.; Rossel, H. Identification and characterization of viruses infecting Bambara groundnut (*Vigna subterranea*) in Nigeria. *Int. J. Pest Manag.* **1997**, *43*, 177–185. [[CrossRef](#)]
109. Kocabas, Z.; Craigon, J.; Azam-Ali, S.N. The germination response of Bambara groundnut (*Vigna subterranea* (L.) Verdc.) to temperature. *Seed Sci. Technol.* **1999**, *27*, 303–313.
110. Sousa, A.C.B.; Carvalho, M.A.; Ramos, A.K.B.; Campos, T.; Sforça, D.A.; Zucchi, M.I.; Jank, L.; Souza, A.P. Genetic studies in *Centrosema pubescens* benth, a tropical forage legume: The mating system, genetic variability and genetic relationships between *Centrosema* species. *Euphytica* **2011**, *181*, 223–235. [[CrossRef](#)]
111. Odu, C.T.I.; Fayemi, A.A.; Ogunwale, J.A. Effect of pH on the growth, nodulation and nitrogen fixation of *Centrosema pubescens* and *Stylosanthes gracilis*. *J. Sci. Food Agric.* **1971**, *22*, 57–59. [[CrossRef](#)]
112. Keller-Grein, G.; Schultze-Kraft, R.; Franco, L.H.; Ramirez, G. Multilocal agronomic evaluation of selected *Centrosema pubescens* germplasm on acid soils. *Trop. Grassl.* **2000**, *34*, 65–77.
113. Morris, J.B. Characterization of butterfly pea (*Clitoria ternatea* L.) accessions for morphology, phenology, reproduction and potential nutraceutical, pharmaceutical trait utilization. *Genet. Resour. Crop. Evol.* **2008**, *56*, 421–427. [[CrossRef](#)]
114. Staples, I.B. *Clitoria ternatea* L. In *Forages, Plant Resources of South-East Asia*; Mannetje, L., Jones, R.M., Eds.; PROSEA Foundation: Bogor, Indonesia, 1992.
115. Adjei-Nsiah, S.; Alabi, B.; Ahiakpa, J.; Kanampiu, F. Response of grain legumes to phosphorus application in the Guinea savanna agro-ecological zones of Ghana. *Agron. J.* **2018**, *110*, 1089–1096. [[CrossRef](#)] [[PubMed](#)]
116. Pueppke, S.G.; Broughton, W.J. *Rhizobium* sp. strain NGR234 and *R. fredii* USDA257 share exceptionally broad, nested host ranges. *Mol. Plant-Microbe Interact.* **1999**, *12*, 293–318. [[CrossRef](#)] [[PubMed](#)]

117. Abaidoo, R.; Dare, M.O.; Killani, S.; Opoku, A. Evaluation of early maturing cowpea (*Vigna unguiculata*) germplasm for variation in phosphorus use efficiency and biological nitrogen fixation potential with indigenous rhizobial populations. *J. Agric. Sci.* **2016**, *155*, 102–116. [CrossRef]
118. Afutu, E.; Agoyi, E.E.; Amayo, R.; Biruma, M.; Rubaihayo, P.R. Cowpea scab disease (*Sphaceloma* sp.) in Uganda. *Crop. Prot.* **2017**, *92*, 213–220. [CrossRef]
119. Ayala-Escobar, V.; Gomez-Jaimes, R.; Santiago-Santiago, V.; Madariaga-Navarrete, A.; Castaneda-Vildoza, A.; Nava-Diaz, C. *Pseudocercospora cruenta* on *Vigna unguiculata* in Mexico. *Australas. Plant Dis. Notes* **2013**, *8*, 115–116. [CrossRef]
120. Chaturvedi, G.S.; Aggarwal, P.K.; Sinha, S.K. Growth and yield of determinate and indeterminate cowpeas in dryland agriculture. *J. Agric. Sci.* **1980**, *94*, 137–144. [CrossRef]
121. Dakora, F.D.; Aboyinga, R.A.; Mahama, Y.; Apaseku, J. Assessment of N₂ fixation in groundnut (*Arachis hypogaea* L.) and cowpea (*Vigna unguiculata* L. Walp) and their relative N contribution to a succeeding maize crop in Northern Ghana. *World J. Microbiol. Biotechnol.* **1987**, *3*, 389–399. [CrossRef]
122. Haruna, I.; Usman, A. Agronomic efficiency of cowpea varieties (*Vigna unguiculata* L. Walp) under varying phosphorus rates in Lafia, Nasarawa state, Nigeria. *Asian J. Crop. Sci.* **2013**, *5*, 209–215. [CrossRef]
123. Wu, X.; Xu, P.; Wang, B.; Lu, Z.; Li, G. Association mapping for *Fusarium* wilt resistance in Chinese asparagus bean germplasm. *Plant Genome* **2015**, *8*, plantgenome2014.11.0082. [CrossRef]
124. Tripathi, K.; Gore, P.G.; Ahlawat, S.P.; Semwal, D.P.; Gautam, N.K.; Kumar, A. Cowpea genetic resources and its utilization: Indian perspective—A review. *Legum. Res. Int. J.* **2019**, *42*, 437–446. [CrossRef]
125. Russi, L.; Acuti, G.; Trabalza-Marinucci, M.; Porta, R.; Rubini, A.; Damiani, F.; Cristiani, S.; Bosco, A.D.; Martuscelli, G.; Bellucci, M.; et al. Genetic characterisation and agronomic and nutritional value of bitter vetch (*Vicia ervilia*), an underutilised species suitable for low-input farming systems. *Crop. Pasture Sci.* **2019**, *70*, 606–614. [CrossRef]
126. Berger, J.; Robertson, L.; Cocks, P. Agricultural potential of Mediterranean grain and forage legumes: Key differences between and within *Vicia* species in terms of phenology, yield, and agronomy give insight into plant adaptation to semi-arid environments. *Genet. Resour. Crop. Evol.* **2002**, *49*, 313–325. [CrossRef]
127. Siddique, K.; Loss, S.P.; Regan, K.L.; Jettner, R.L. Adaptation and seed yield of cool season grain legumes in Mediterranean environments of south-western Australia. *Aust. J. Agric. Res.* **1999**, *50*, 375. [CrossRef]
128. Larbi, A.; El-Moneim, A.A.; Nakkoul, H.; Jammal, B.; Hassan, S. Intra-species variations in yield and quality determinants in *Vicia* species: 1. Bitter vetch (*Vicia ervilia* L.). *Anim. Feed. Sci. Technol.* **2011**, *165*, 278–287. [CrossRef]
129. Saoub, H.M.; Akash, M.W. Variations among two vetch landrace species in Jordan. *J. Food Agric. Environ.* **2012**, *10*, 763–767.
130. Government of Saskatchewan Inoculation of Pulse Crops. Soils, Fertility, and Nutrients. 2020. Available online: <https://www.saskatchewan.ca/business/agriculture-natural-resources-and-industry/agribusiness-farmers-and-ranchers/crops-and-irrigation/soils-fertility-and-nutrients/inoculation-of-pulse-crops> (accessed on 29 November 2018).
131. Branch, S. Fenugreek (*Trigonella foenum-graecum* L.) as a valuable medicinal plant. *Int. J. Adv. Biol. Biomed. Res.* **2013**, *1*, 922–931.
132. Baldoni, R.; Giardini, L. *Coltivazioni Erbacee Cereali e Proteaginose*; Patron Editore: Granarolo dell’Emilia, Italy, 2001; ISBN 978-88-555-2541-1.
133. Patto, M.V.; Rubiales, D. Resistance to rust and powdery mildew in *Lathyrus* crops. *Czech J. Genet. Plant Breed.* **2014**, *50*, 116–122. [CrossRef]
134. Niembro Rocas, A. *Enterolobium cyclocarpum* (Jacq.) Griseb. In *Tropical Tree Seed Manual; Agricultural Handbook*; Vozzo, J.A., Ed.; USDA Forest Service: Washington, DC, USA, 2002; pp. 449–451.
135. Krishna, K.R. *Agroecosystems of South India Nutrient Dynamics, Ecology and Productivity*; Brown Walker Press: Boca Raton, FL, USA, 2010; ISBN 1-59942-533-5.
136. Jansen, P.C.M. *Macrotyloma uniflorum* (Lam.) Verdc. In *Pulses; Plant Resources of South-East, Asia*; van der Maesen, L.J.G., Somaatmadja, S., Eds.; PUDOC Publishing: Bogor, Indonesia, 1989.
137. Mohammed, M.; Sowley, E.; Dakora, F. Variations in N₂ fixation of field-grown Kersting’s groundnut (*Macrotyloma geocarpum*) landraces in response to inoculation with two *Bradyrhizobium* strains in the northern region of Ghana. *S. Afr. J. Bot.* **2016**, *103*, 333. [CrossRef]
138. PROTA4U *Macrotyloma geocarpum* (Harms) Marechal. 2020. Available online: [https://www.prota4u.org/database/protav8.asp?fr=1&g=pe&p=Macrotyloma+geocarpum+\(Harms\)+Mar%E9chal+&+Baudet](https://www.prota4u.org/database/protav8.asp?fr=1&g=pe&p=Macrotyloma+geocarpum+(Harms)+Mar%E9chal+&+Baudet) (accessed on 29 November 2018).
139. Université de Liège. *Fiche Technique du Mucuna*; Université de Liège: Liège, Belgium, 2014; Volume 1.
140. Sheahan, C. *Plant Guide for Jack Bean (Canavalia ensiformis)*; USDA Natural Resources Conservation Service: Washington, DC, USA, 2012; Volume 4.
141. Miamoto, A.; Dias-Arieira, C.R.; Cardoso, M.R.; Puerari, H.H. Penetration and reproduction of *Meloidogyne javanica* on leguminous crops. *J. Phytopathol.* **2016**, *164*, 890–895. [CrossRef]
142. Roy, S.S.; Kumar, S.; Sharma, S.; Devi, A.; Singh, N.; Prakash, N.; Ngachan, S. Tree bean (*Parkia roxburghii*): A potential multipurpose tree legume of northeast India. In *Proceedings of the Souvenir & Abstracts National Symposium on Vegetable Legumes for soil and Human Health*, Varanasi, India, 12 February 2016; pp. 201–208.
143. Long, R.; Temple, S.; Schmierer, J.; Canevari, M.; Meyer, R.D. *Common Dry Bean Production in California*, 2nd ed.; University of California Press: Oakland, CA, USA, 2010.

144. Bulyaba, R.; Lenssen, A.W. Influence of *Bradyrhizobium* inoculation and fungicide seed treatment on development and yield of cowpea, lablab, and soybean. *Crop. Forage Turfgrass Manag.* **2017**, *3*, 1–7. [[CrossRef](#)]
145. Díaz, M.F.; González, A.; Padilla, C.; Curbelo, F. Performance of forage and grain production of *Canavalia ensiformis*, *Lablab purpureus* and *Stizolobium niveum* plantations sown in September. *Cuba. J. Agric. Sci.* **2003**, *37*, 65–71.
146. Rangaiah, D.V.; Dsouza, M. Hyacinth bean (*Lablab purpureus*): An adept adaptor to adverse environments. *Legume Perspectives* **2016**, *13*, 20.
147. Long, R.; Temple, S.; Meyer, R.; Schwankl, L.; Godfrey, L.; Canevari, M.; Roberts, P. *Lima Bean Production in California*; University of California, Agriculture and Natural Resources: Berkley, CA, USA, 2014; ISBN 978-1-60107-860-5.
148. De Araújo, A.S.F.; Antunes, J.E.L.; de Almeida Lopes, A.C.; Ferreira Gomes, R.L. Rhizobia and Lima Bean Symbiosis: Importance, Occurrence and Diversity. 2015. Available online: https://www.researchgate.net/publication/297766907_Rhizobia_and_lima_bean_symbiosis_importance_occurrenceand_diversity (accessed on 27 November 2018).
149. Marsh, L.E.; Davis, D.W. Influence of high temperature on the performance of some *Phaseolus* species at different developmental stages. *Euphytica* **1985**, *34*, 431–439. [[CrossRef](#)]
150. Brink, M.; Belay, G. *Plant Resources of Tropical Africa 1. Cereals And Pulses*; PROTA Foundation: Wageningen, The Netherlands; Backhuys Publishers: Leiden, The Netherlands; CTA: Wageningen, The Netherlands, 2006; ISBN 978-90-5782-170-7.
151. Pooniya, V.; Choudhary, A.K.; Dass, A.; Bana, R.S.; Rana, K.S.; Rana, D.S.; Tyagi, V.K.; Puniya, M.M. Improved crop management practices for sustainable pulse production: An Indian perspective. *Indian J. Agric. Sci.* **2015**, *85*, 747–758.
152. Van Oers, C.C.C.M. Vigna aconitifolia (Jacq.) Maréchal. In *Pulses, Plant Resources of South-East Asia*; van der Maesen, L.J.G., Somaatmadja, S., Eds.; PUDOC Publishing: Bogor, Indonesia, 1989.
153. Riaz Malik, S.; Imran, M.; Asadullah, M.; Jawad, M.; Sarwar, A. Pulses Program. 2018. Available online: <http://www.parc.gov.pk/index.php/en/csi/137-narc/crop-sciences-institut/712-national-coordinated-pulses-programme> (accessed on 29 November 2018).
154. Tariq, S.; Ali, S.; Ijaz, S.S. Improving nitrogen fixation capacity and yield of mungbean and mashbean by phosphorous management in Pothowar. *Sarhad J. Agric.* **2007**, *23*, 6.
155. Chinnasamy, G.; Bal, A.; McKenzie, D. Fatty acid composition of grass pea (*Lathyrus sativus* L.) seeds. *Lathyrus Lathyrism Newsl.* **2005**, *4*, 2–4.
156. Akhter, M.; Akanda, A.; Kobayashi, K.; Jain, R.; Mandal, B. Plant virus diseases and their management in Bangladesh. *Crop. Prot.* **2019**, *118*, 57–65. [[CrossRef](#)]
157. Hardwick, R.C. Review of recent research on navy beans (*Phaseolus vulgaris*) in the United Kingdom. *Ann. Appl. Biol.* **1988**, *113*, 205–227. [[CrossRef](#)]
158. Gonzalez, D.; Obrador, A.A.; Alvarez, J.M. Behavior of zinc from six organic fertilizers applied to a navy bean crop grown in a calcareous soil. *J. Agric. Food Chem.* **2007**, *55*, 7084–7092. [[CrossRef](#)] [[PubMed](#)]
159. Siddique, K.; Loss, S.; Enneking, D. Narbon bean (*Vicia narbonensis* L.): A promising grain legume for low rainfall areas of south-western Australia. *Aust. J. Exp. Agric.* **1996**, *36*, 53–62. [[CrossRef](#)]
160. Nadal, S.; Moreno, M.T. Behaviour of Narbon bean (*Vicia narbonensis* L.) under presence-absence of broomrape (*Orobanche crenata* Forsk.) in rainfed agricultural systems in southern Spain. *J. Sustain. Agric.* **2007**, *30*, 133–143. [[CrossRef](#)]
161. Enneking, D.; Maxted, N. Narbon bean (*Vicia narbonensis* L.). In *Evolution of Crop Plants*, 2nd ed.; Smartt, J., Simmonds, N.W., Eds.; Longman: London, UK, 1995; pp. 316–321.
162. Rao, J.V.D.K.K.; Dart, P.J. Nodulation, nitrogen fixation and nitrogen uptake in pigeonpea (*Cajanus cajan* (L.) Millsp) of different maturity groups. *Plant Soil* **1987**, *99*, 255–266. [[CrossRef](#)]
163. Smartt, J. Evolution of grain legumes. II. Old and new world pulses of lesser economic importance. *Exp. Agric.* **1985**, *21*, 1–18. [[CrossRef](#)]
164. Ahlawat, I.P.S.; Gangaiah, B.; Singh, I.P. Pigeonpea (*Cajanus cajan*) research in India—An overview. *Indian J. Agric. Sci.* **2005**, *75*, 309–320.
165. Osorno, J.M.; Wal, A.J.V.; Kloberdanz, M.; Pasche, J.S.; Schroder, S.; Miklas, P.N. A new slow-darkening Pinto bean with improved agronomic performance: Registration of “ND-Palomino”. *J. Plant Regist.* **2017**, *12*, 25–30. [[CrossRef](#)]
166. Osorno, J.M.; Grafton, K.F.; Rojas-Cifuentes, G.A.; Gelin, R.; Wal, A.J.V. Registration of “Lariat” and “Stampede” Pinto beans. *J. Plant Regist.* **2010**, *4*, 5–11. [[CrossRef](#)]
167. Adjolohoun, S.; Bindelle, J.; Adandedjan, C.C.; Tolebq, S.S.; Houinato, M.R.; Sinsin, A.B. Variety and environmental effects on crude protein concentration and mineral composition of *Arachis pintoi* (Kaprovicak & Gregory) in Benin (West Africa). *J. Appl. Biol. Biotechnol.* **2013**, *1*, 24–28. [[CrossRef](#)]
168. Heuzé, V.; Tran, G.; Delagarde, R.; Bastianelli, D.; Lebas, F. *Pinto peanut (Arachis pintoi)*; INRA: Paris, France; CIRAD: Montpellier, France; AFZ: Ajman, United Arab Emirates; FAO: Rome, Italy, 2016.
169. Cook, B.G. *Arachis pintoi* Krap. & Greg., Nom. Nud. In *Forages; Plant Resources of South-East, Asia*; Mannetje, L., Jones, R.M., Eds.; PUDOC Publishing: Bogor, Indonesia, 1992.
170. Doignon-Bourcier, F.; Sy, A.; Willems, A.; Torck, U.; Dreyfus, B.; Gillis, M.; de Lajudie, P. Diversity of *Bradyrhizobia* from 27 tropical leguminosae species native of Senegal. *Syst. Appl. Microbiol.* **1999**, *22*, 647–661. [[CrossRef](#)]
171. Khadka, K.; Acharya, B.D. *Cultivation Practices of Ricebean*; Local Initiatives for Biodiversity, Research and Development (LI-BIRD): Pokhara, Nepal, 2009; 31p, ISBN 978-9937-8145-1-5.

172. Dhillon, P.K.; Tanwar, B. Rice bean: A healthy and cost-effective alternative for crop and food diversity. *Food Secur.* **2018**, *10*, 525–535. [CrossRef]
173. Kapoor, C.; Gopi, R.; Karuppaiyan, R. Ricebean: An underutilized pulse crop of Sikkim. *Asian Agri-Hist.* **2012**, *16*, 417–421.
174. Pattanayak, A.; Roy, S.; Sood, S.; Iangrai, B.; Banerjee, A.; Gupta, S.; Joshi, D.C. Rice bean: A lesser-known pulse with well-recognized potential. *Planta* **2019**, *250*, 873–890. [CrossRef] [PubMed]
175. Kimani, P.M.; Njau, S.; Mulanya, M.; Narla, R.D. Breeding runner bean for short-day adaptation, grain yield, and disease resistance in eastern Africa. *Food Energy Secur.* **2019**, *8*, e171. [CrossRef]
176. Sileshi, G.; Maghembe, J.; Rao, M.; Ogol, C.; Sithanatham, S. Insects feeding on *Sesbania* species in natural stands and agroforestry systems in southern Malawi. *Agrofor. Syst.* **2000**, *49*, 41–52. [CrossRef]
177. Sileshi, G.; Mafongoya, P. Incidence of *Mesoplatys ochroptera* Stål (Coleoptera: Chrysomelidae) on *Sesbania sesban* in pure and mixed species fallows in eastern Zambia. *Agrofor. Syst.* **2002**, *56*, 225–231. [CrossRef]
178. Heering, J.H.; Guteridge, R.C. *Sesbania sesban* (L.) Merrill. In *Forages; Plant Resources of South-East, Asia*; Mannerje, L., Jones, R.M., Eds.; PUDOC Publishing: Bogor, Indonesia, 1992.
179. Yusuf, A.; Mofio, B.; Ahmed, A. Proximate and mineral composition of *Tamarindus indica* linn 1753 seeds. *Sci. World J.* **2010**, *2*. [CrossRef]
180. El-Siddig, K.; Gunasena, H.P.M.; Prasad, B.A.; Pushpakumara, D.K.N.G.; Ramana, K.V.R.; Vijayanand, P.; Williams, J.T. Tamarind: *Tamarindus indica* L. In *Fruits for the Future 1*; Williams, J.T., Smith, R.W., Haq, Z., Eds.; Southampton Centre for Underutilised Crops: Southampton, UK, 2006.
181. Bowe, C.; Haq, N. Quantifying the global environmental niche of an underutilised tropical fruit tree (*Tamarindus indica*) using herbarium records. *Agric. Ecosyst. Environ.* **2010**, *139*, 51–58. [CrossRef]
182. Cardet, C.; Kandji, T.; Delobel, A.; Danthu, P. Efficiency of neem and groundnut oils in protecting leguminous tree seeds against seed beetles in the Sahel. *Agrofor. Syst.* **1998**, *40*, 29–40. [CrossRef]
183. Castellano, G.; Lugo, Z.; Casassa-Padrón, A.M.; Pérez-Pérez, E.; Núñez-Castellano, K. Plant parasitic nematodes associated to potentials fruit trees, in three areas of Mara, Zulia state, Venezuela. *Rev. Fac. Agron.* **2014**, *31*, 414–422.
184. Urrutia Gutierrez, W. *Determinacion de Parametros Optimos de Extraccion Alcalina para la Obtencion de Aislado Proteico a Partir de Tarwi (Lupinus Mutabilis)*; Universidad Nacional Micaela Bastidas de Apurímac: Abancay, Peru, 2010.
185. Mogotsi, K.K. *Phaseolus acutifolius*. 2018. Available online: <https://www.prota4u.org/database/protav8.asp?fr=1&g=pe&p=Phaseolus+acutifolius+A.Gray> (accessed on 28 November 2018).
186. Mishra, S.; Macedo, M.; Panda, S.; Panigrahi, J. Bruchid pest management in pulses: Past practices, present status and use of modern breeding tools for development of resistant varieties. *Ann. Appl. Biol.* **2017**, *172*, 4–19. [CrossRef]
187. Aeron, A.; Maheshwari, D.K.; Dheeman, S.; Agarwal, M.; Dubey, R.; Bajpai, V.K. Plant growth promotion and suppression of charcoal-rot fungus (*Macrophomina phaseolina*) in velvet bean (*Mucuna pruriens* L.) by root nodule bacteria. *J. Phytopathol.* **2017**, *165*, 463–478. [CrossRef]
188. Arora, N.K.; Kang, S.C.; Maheshwari, D.K. Isolation of siderophore-producing strains of *Rhizobium meliloti* and their biocontrol potential against *Macrophomina phaseolina* that causes charcoal rot of groundnut. *Curr. Sci.* **2001**, *81*, 673–677.
189. Baiswar, P.; Mohapatra, K.P.; Chandra, P.; Ngachan, S.V. Rust resistance in *Mucuna pruriens* accessions. *Indian Phytopathol.* **2017**, *70*, 498–499. [CrossRef]
190. Wuljarni-Soetjipto, N.; Maligali, R.F. *Mucuna pruriens* (L.) DC. Cv. group Utilis. In *Auxiliary Plants. Plant Resources of South-East Asia*; Faridah Hanum, I., van der Maesen, L.J.G., Eds.; PROSEA Foundation: Bogor, Indonesia, 1997.
191. Okito, A.; Alves, B.J.R.; Urquiaga, S.; Boddey, R. Nitrogen fixation by groundnut and velvet bean and residual benefit to a subsequent maize crop. *Fertilization* **2004**, *39*, 1183–1190. [CrossRef]
192. National Academies. *The Winged Bean: A High Protein Crop for the Tropics. Report of an Ad Hoc Panel of the Advisory Committee on Technology Innovation Board on Science and Technology for International Development*; National Academies: Washington, DC, USA, 1975.
193. Sfrensen, M.; van Hoof, W.C.H. *Pachyrhizus erosus* (L.) urban. In *Plants Yielding Non-Seed Carbohydrates. Plant Resources of South-East Asia*; Flach, M., Rumawas, F., Eds.; PROSEA Foundation: Bogor, Indonesia, 1996.
194. Susan John, K.; George, J.; Shanida Beegum, S.U.; Shivay, Y.S. Soil fertility and nutrient management in tropical tuber crops—An overview. *Indian J. Agron.* **2016**, *61*, 263–273.
195. Unkovich, M.; Pate, J.; Armstrong, E.; Sanford, P. Nitrogen economy of annual crop and pasture legumes in southwest Australia. *Soil Biol. Biochem.* **1995**, *27*, 585–588. [CrossRef]
196. Temesgen, T.; Keneni, G.; Sefera, T.; Jarso, M. Yield stability and relationships among stability parameters in Faba bean (*Vicia faba* L.) genotypes. *Crop. J.* **2015**, *3*, 258–268. [CrossRef]
197. Ahmed, K.; Awan, M.S. Integrated management of insect pests of chickpea *Cicer arietinum* (L. Walp) in south Asian countries: Present status and future strategies—A review. *Pak. J. Zool.* **2013**, *45*, 1125–1145.
198. Tiwari, K.; Dwivedi, B.; Pathak, A. Evaluation of iron pyrites as Sulphur fertilizer. *Nutr. Cycl. Agroecosyst.* **1984**, *5*, 235–243. [CrossRef]
199. Thangwana, N.M.; Ogola, J.B.O. Yield and yield components of chickpea (*Cicer arietinum*): Response to genotype and planting density in summer and winter sowings. *J. Food Agric. Environ.* **2012**, *10*, 710–715.
200. Maqbool, A.; Shafiq, S.; Lake, L. Radiant frost tolerance in pulse crops—A review. *Euphytica* **2009**, *172*, 1–12. [CrossRef]

201. Mola, T.; Alemayehu, S.; Fikre, A.; Ojiewo, C.; Alemu, K.; Abdi, T. Heat tolerance responses of chickpea (*Cicer arietinum* L.) genotypes in the thermal zone of Ethiopia, a case of Werer station. *Ethiop. J. Crop Sci.* **2018**, *6*, 95–118.
202. Mukoko, O.; Galwey, N.; Allen, D. Developing cultivars of the common bean (*Phaseolus vulgaris* L.) for southern Africa: Bean common mosaic virus resistance, consumer preferences and agronomic requirements. *Field Crop. Res.* **1995**, *40*, 165–177. [[CrossRef](#)]
203. Tigist, S.G.; Melis, R.; Sibiya, J.; Keneni, G. Evaluation of different Ethiopian common bean, *Phaseolus vulgaris* (Fabaceae) genotypes for host resistance to the Mexican bean weevil, *Zabrotes subfasciatus* (Coleoptera: Bruchidae). *Int. J. Trop. Insect Sci.* **2017**, *38*, 1–15. [[CrossRef](#)]
204. Tigist, S.G.; Melis, R.; Sibiya, J.; Amelework, A.B.; Keneni, G.; Tegene, A. Population structure and genome-wide association analysis of bruchid resistance in Ethiopian common bean genotypes. *Crop. Sci.* **2019**, *59*, 1504–1515. [[CrossRef](#)]
205. Swegarden, H.R.; Sheaffer, C.C.; Michaels, T.E. Yield stability of heirloom dry bean (*Phaseolus vulgaris* L.) cultivars in midwest organic production. *HortScience* **2016**, *51*, 8–14. [[CrossRef](#)]
206. Soratto, R.P.; Perez, A.A.G.; Fernandes, A.M. Age of no-till system and nitrogen management on common bean nutrition and yield. *Agron. J.* **2014**, *106*, 809–820. [[CrossRef](#)]
207. Maingi, J.M.; Shisanya, C.A.; Gitonga, N.M.; Hornetz, B. Nitrogen fixation by common bean (*Phaseolus vulgaris* L.) in pure and mixed stands in semi-arid southeast Kenya. *Eur. J. Agron.* **2001**, *14*, 1–12. [[CrossRef](#)]
208. Hungria, M.; Campo, R.J.; Mendes, I. Benefits of inoculation of the common bean (*Phaseolus vulgaris*) crop with efficient and competitive *Rhizobium tropici* strains. *Biol. Fertil. Soils* **2003**, *39*, 88–93. [[CrossRef](#)]
209. Hernandez-Armenta, R.; Wien, H.C.; Eaglesham, A.R.J. Carbohydrate partitioning and nodule function in common bean after heat stress. *Crop. Sci.* **1989**, *29*, 1292–1297. [[CrossRef](#)]
210. Hernández, G.; Ramírez, M.; Valdés-López, O.; Tesfaye, M.; Graham, M.A.; Czechowski, T.; Schlereth, A.; Wandrey, M.; Erban, A.; Cheung, F.; et al. Phosphorus stress in common bean: Root transcript and metabolic responses. *Plant Physiol.* **2007**, *144*, 752–767. [[CrossRef](#)] [[PubMed](#)]
211. Gutiérrez, A.; Mariot, E.; Cure, J.; Riddle, C.; Ellis, C.; Villacorta, A. A model of bean (*Phaseolus vulgaris* L.) growth types I–III: Factors affecting yield. *Agric. Syst.* **1994**, *44*, 35–63. [[CrossRef](#)]
212. Graham, P.; Ranalli, P. Common bean (*Phaseolus vulgaris* L.). *Field Crop. Res.* **1997**, *53*, 131–146. [[CrossRef](#)]
213. Głowacka, A.; Gruszecki, T.; Szostak, B.; Michałek, S. The response of common bean to sulphur and Molybdenum fertilization. *Int. J. Agron.* **2019**, *2019*, 3830712. [[CrossRef](#)]
214. Tedford, E.C.; Inglis, D.A. Evaluation of legumes common to the pacific northwest as hosts for the pea cyst nematode, *Heterodera goettingiana*. *J. Nematol.* **1999**, *31*, 155–163.
215. Khan, Z.; Gautam, N.K.; Gawade, B.H.; Dubey, S.C. Screening of lentil (*Lens culinaris* Medik.) germplasm for resistance to root-knot nematode, *Meloidogyne incognita*. *Indian J. Genet. Plant Breed.* **2017**, *77*, 408. [[CrossRef](#)]
216. Huang, J.; Afshar, R.K.; Chen, C. Lentil response to nitrogen application and Rhizobia inoculation. *Commun. Soil Sci. Plant Anal.* **2016**, *47*, 2458–2464. [[CrossRef](#)]
217. Clark, S. *Plant Guide for White Lupine* (*Lupinus albus* L.); USDA-NRCS Big Flats Plant Materials Center: Corning, NY, USA, 2014.
218. Wylie, S.; Wilson, C.; Jones, R.; Jones, M. A polymerase chain reaction assay for cucumber mosaic virus in lupin seeds. *Aust. J. Agric. Res.* **1993**, *44*, 41–51. [[CrossRef](#)]
219. Unkovich, M.; Pate, J.; Hamblin, J. The nitrogen economy of broadacre lupin in southwest Australia. *Aust. J. Agric. Res.* **1994**, *45*, 149–164. [[CrossRef](#)]
220. Tang, C.; Zheng, S.J.; Qiao, Y.F.; Wang, G.H.; Han, X.Z. Interactions between high pH and iron supply on nodulation and iron nutrition of *Lupinus albus* L. genotypes differing in sensitivity to iron deficiency. *Plant Soil* **2006**, *279*, 153–162. [[CrossRef](#)]
221. Vijayakumari, K.; Siddhuraju, P.; Janardhanan, K. Nutritional assessment and chemical composition of the lesser-known tree legume, *Acacia leucophloea* (Roxb.) Willd. *Food Chem.* **1994**, *50*, 285–288. [[CrossRef](#)]
222. Yoshida, K.; Sato, H.; Sato, M. The extent and its source of variation for characteristics related to seed quality of Adzuki beans. III. The water uptake of seeds and hard seededness. *Jpn. J. Crop. Sci.* **1995**, *64*, 7–13. [[CrossRef](#)]
223. Koura, K.; Ouidoh, P.; Azokpota, P.; Ganglo, J.C.; Hounhouigan, D.J. Caractérisation physique et composition chimique des graines de *Parkia biglobosa* (Jacq.) R. Br. en usage au Nord-Bénin. *J. Appl. Biosci.* **2014**, *75*, 6239. [[CrossRef](#)]
224. Gernmah, D.I.; Atolagbe, M.O.; Echegwo, C.C. Nutritional composition of the African locust bean (*Parkia biglobosa*) fruit pulp. *Niger. Food J.* **2007**, *25*, 190–196. [[CrossRef](#)]
225. Abdulrahman, B.O.; Osibemhe, M.; Idoko, A.S. The status of mineral and anti-nutritional composition of raw and fermented seeds of African locust bean (*Parkia biglobosa*). *Int. J. Curr. Res. Biosci. Plant Biol.* **2016**, *3*, 1–4. [[CrossRef](#)]
226. Enujiugha, V.N.; Ayodele-Oni, O. Evaluation of nutrients and some anti-nutrients in lesser-known, underutilized oilseeds. *Int. J. Food Sci. Technol.* **2003**, *38*, 525–528. [[CrossRef](#)]
227. Ifesan, B.O.T.; Akintade, A.O.; Gabriel-Ajobiwe, R.A.O. Physicochemical and nutritional properties of *Mucuna pruriens* and *Parkia biglobosa* subjected to controlled fermentation. *Int. Food Res. J.* **2017**, *24*, 2177–2184.
228. Ikhuoria, E.U. Characteristics and composition of African oil bean seed (*Pentaclethra macrophylla* Benth). *J. Appl. Sci.* **2008**, *8*, 1337–1339. [[CrossRef](#)]
229. Adeyeye, E.I.; Oshodi, A.A.; Ipinmoroti, K.O. Functional properties of some varieties of African yam bean (*Sphenostylis stenocarpa*) flour II. *Int. J. Food Sci. Nutr.* **1994**, *45*, 115–126. [[CrossRef](#)]

230. Ajayi, F.T.; Akande, S.R.; Odejide, J.O.; Idowu, B. Nutritive evaluation of some tropical under-utilized grain legume seeds for ruminant's nutrition. *J. Am. Sci.* **2010**, *7*, 1–7.
231. Leung, W.; Busson, F.; Jardin, C. *Food Composition Table for Use in Africa*; FAO: Rome, Italy, 1968.
232. Ijarotimi, O.S.; Esho, T.R. Comparison of nutritional composition and anti-nutrient status of fermented, germinated and roasted bambara groundnut seeds (*Vigna subterranea*). *Br. Food J.* **2009**, *111*, 376–386. [[CrossRef](#)]
233. Akubuo, C.O.; Uguru, M.I. Studies on the nutritive characteristics and fracture resistance to compressive loading of selected Bambara groundnut lines. *J. Sci. Food Agric.* **1999**, *79*, 2063–2066. [[CrossRef](#)]
234. Amarteifio, J.O.; Karikari, S.K.; Modise, O.J. The proximate and mineral composition of six landraces of Bambara groundnut. *Trop. Sci.* **2002**, *42*, 188–191.
235. Halimi, R.A.; Barkla, B.J.; Mayes, S.; King, G.J. The potential of the underutilized pulse Bambara groundnut (*Vigna subterranea* (L.) Verdc.) for nutritional food security. *J. Food Compos. Anal.* **2018**, *77*, 47–59. [[CrossRef](#)]
236. Iyayi, E.A.; Kluth, H.; Rodehutsord, M. Chemical composition, antinutritional constituents, precaecal crude protein and amino acid digestibility in three unconventional tropical legumes in broilers. *J. Sci. Food Agric.* **2006**, *86*, 2166–2171. [[CrossRef](#)]
237. Baptista, A.; Pinho, O.; Pinto, E.; Casal, S.; Mota, C.; Ferreira, I.M. Characterization of protein and fat composition of seeds from common beans (*Phaseolus vulgaris* L.), cowpea (*Vigna unguiculata* L. Walp) and Bambara groundnuts (*Vigna subterranea* L. Verdc) from Mozambique. *J. Food Meas. Charact.* **2016**, *11*, 442–450. [[CrossRef](#)]
238. Da Silva, D.O.M.; Santos, C.A.F.; Boiteux, L.S. Adaptability and stability parameters of total seed yield and protein content in cowpea (*Vigna unguiculata*) genotypes subjected to semi-arid conditions. *Aust. J. Crop Sci.* **2016**, *10*, 1164–1169. [[CrossRef](#)]
239. Berger, J.D.; Siddique, K.H.M.; Loss, S.P. Cool season grain legumes for Mediterranean environments: Species × environment interaction in seed quality traits and anti-nutritional factors in the genus *Vicia*. *Aust. J. Agric. Res.* **1999**, *50*, 389. [[CrossRef](#)]
240. Bakoglu, A.; Bagci, E.; Ciftci, H. Fatty acids, protein contents and metal composition of some feed crops from Turkey. *J. Food Agric. Environ.* **2009**, *7*, 343–346.
241. Martín-Pedrosa, M.; Varela, A.; Guillamón, E.; Cabellos, B.; Burbano, C.; Gomez-Fernandez, J.; de Mercado, E.; Gomez-Izquierdo, E.; Cuadrado, C.; Múzquiz, M. Biochemical characterization of legume seeds as ingredients in animal feed. *Span. J. Agric. Res.* **2016**, *14*, e0901. [[CrossRef](#)]
242. Sadeghi, G.; Mohammadi, L.; Ibrahim, S.; Gruber, K. Use of bitter vetch (*Vicia ervilia*) as a feed ingredient for poultry. *World's Poult. Sci. J.* **2009**, *65*, 51. [[CrossRef](#)]
243. Pastor-Cavada, E.; Juan, R.; Pastor, J.E.; Alaiz, M.; Vioque, J. Nutritional characteristics of seed proteins in 28 *Vicia* species (Fabaceae) from southern Spain. *J. Food Sci.* **2011**, *76*, C1118–C1124. [[CrossRef](#)] [[PubMed](#)]
244. Wani, S.A.; Kumar, P. Fenugreek: A review on its nutraceutical properties and utilization in various food products. *J. Saudi Soc. Agric. Sci.* **2018**, *17*, 97–106. [[CrossRef](#)]
245. Abou-Shleel, S. Effect of air temperature on growth, yield and active ingredients of Fenugreek (*Trigonella foenum-graecum*). *Nat. Sci.* **2014**, *12*, 50–54.
246. Hanbury, C.; White, C.; Mullan, B.; Siddique, K. A review of the potential of *Lathyrus sativus* L. and *L. cicera* L. grain for use as animal feed. *Anim. Feed. Sci. Technol.* **2000**, *87*, 1–27. [[CrossRef](#)]
247. Folarin, O.M.; Igbon, O.C. Chemical composition of *Enterolobium cyclocarpum* (Jacq.) Griseb. seed and physicochemical properties of the oil extract. *Hamdard Med.* **2010**, *53*, 21–26.
248. Ravindran, R.; Sundar, S.T.B. Nutritive value of horse gram (*Dolichos biflorus*) for egg type chicks and growers. *Tamilnadu J. Vet. Anim. Sci.* **2009**, *5*, 125–131.
249. Bhartiya, A.; Aditya, J.P.; Kant, L. Nutritional and remedial potential of an underutilized food legume horse gram (*Macrotyloma uniflorum*): A review. *J. Anim. Plant Sci.* **2015**, *25*, 908–920.
250. Prasad, S.K.; Singh, M.K. Horse gram—An underutilized nutraceutical pulse crop: A review. *J. Food Sci. Technol.* **2014**, *52*, 2489–2499. [[CrossRef](#)]
251. Kalidass, C.; Mohan, V. Nutritional and antinutritional composition of itching bean (*Mucuna pruriens* (L.) DC Var. *pruriens*): An underutilized tribal pulse in western Ghats, Tamil Nadu. *Trop. Subtrop. Agroecosyst.* **2011**, *14*, 279–293.
252. Adebowale, Y.A.; Adeyemi, I.A.; Oshodi, A.A. Functional and physicochemical properties of flours of six *Mucuna* species. *Afr. J. Biotechnol.* **2005**, *4*, 1461–1468.
253. Abitogun, A.S.; Olasehinde, E.F. Nutritional evaluation of seed and characterization of crude Jack bean (*Canavalia ensiformis*) oil. *IOSR J. Appl. Chem.* **2012**, *1*, 36–40. [[CrossRef](#)]
254. Agbede, J.O. Characterisation of the leaf meals, protein concentrates and residues from some tropical leguminous plants. *J. Sci. Food Agric.* **2006**, *86*, 1292–1297. [[CrossRef](#)]
255. Salam, J.; Singh, P.K.; Dutta, B.K.; Sahoo, U. Chemical composition and nutritive indices in *Parkia roxburghii* g. Don, a leguminous plant of India. *Indian J. Agric. Biochem.* **2009**, *22*, 87–93.
256. Longvah, T.; Deosthale, Y. Nutrient composition and food potential of *Parkia roxburghii*, a less known tree legume from northeast India. *Food Chem.* **1998**, *62*, 477–481. [[CrossRef](#)]
257. Blair, M.W.; Porch, T.; Cichy, K.; Galeano, C.H.; Lariguet, P.; Pankhurst, C.; Broughton, W. Induced mutants in common bean (*Phaseolus vulgaris*), and their potential use in nutrition quality breeding and gene discovery. *Isr. J. Plant Sci.* **2007**, *55*, 191–200. [[CrossRef](#)]

258. Gwanzura, T.; Ng'ambi, J.; Norris, D. Nutrient composition and tannin contents of forage Sorghum, cowpea, lablab and Mucuna hays grown in Limpopo Province of south Africa. *Asian J. Anim. Sci.* **2012**, *6*, 256–262. [[CrossRef](#)]
259. Bhardwaj, H.L.; Hamama, A.A. Genotype and environment effects on lablab seed yield and composition. *HortScience* **2019**, *54*, 2156–2158. [[CrossRef](#)]
260. Powell, A.M. Marama bean (*Tylosema esculentum*, Fabaceae) seed crop in Texas. *Econ. Bot.* **1987**, *41*, 216–220. [[CrossRef](#)]
261. Holse, M.; Husted, S.; Hansen, S. Chemical composition of Marama bean (*Tylosema esculentum*)—A wild African bean with unexploited potential. *J. Food Compos. Anal.* **2010**, *23*, 648–657. [[CrossRef](#)]
262. Amarteifio, J.; Moholo, D. The chemical composition of four legumes consumed in Botswana. *J. Food Compos. Anal.* **1998**, *11*, 329–332. [[CrossRef](#)]
263. Gunathilake, K.T.; Herath, T.; Wansapala, J. Comparison of physicochemical properties of selected locally available legume varieties (mung bean, cowpea and soybean). *Potravin. Slovak J. Food Sci.* **2016**, *10*, 424–430. [[CrossRef](#)]
264. Hadjipanayiotou, M.; Economides, S. Chemical composition, in situ degradability and amino acid composition of protein supplements fed to livestock and poultry in Cyprus. *Livest. Res. Rural. Dev.* **2001**, *13*, 56.
265. Brand, T.; Brandt, D.; Cruywagen, C. Chemical composition, true metabolisable energy content and amino acid availability of grain legumes for poultry. *S. Afr. J. Anim. Sci.* **2004**, *34*, 116–122. [[CrossRef](#)]
266. Anton, A.A.; Ross, K.A.; Beta, T.; Fulcher, R.G.; Arntfield, S.D. Effect of pre-dehulling treatments on some nutritional and physical properties of navy and pinto beans (*Phaseolus vulgaris* L.). *LWT* **2008**, *41*, 771–778. [[CrossRef](#)]
267. Grosso, N.R.; Nepote, V.; Guzmán, C.A. Chemical composition of some wild peanut species (*Arachis* L.) seeds. *J. Agric. Food Chem.* **2000**, *48*, 806–809. [[CrossRef](#)]
268. Fathima, K.R.; Mohan, V. Nutritional and antinutritional assessment of *Mucuna atropurpurea* DC: An underutilized tribal pulse. *Afr. J. Basic Appl. Sci.* **2009**, *1*, 129–136.
269. Siddhuraju, P.; Vijayakumari, K.; Janardhanan, K. The biochemical composition and nutritional potential of the tribal pulse, *Alysicarpus rugosus* (Willd.) DC. *Food Chem.* **1992**, *45*, 251–255. [[CrossRef](#)]
270. Kallah, M.; Bale, J.; Abdullahi, U.; Muhammad, I.; Lawal, R. Nutrient composition of native forbs of semi-arid and dry sub-humid savannas of Nigeria. *Anim. Feed. Sci. Technol.* **2000**, *84*, 137–145. [[CrossRef](#)]
271. Nigussie, Z.; Alemayehu, G. *Sesbania sesban* (L.) Merrill: Potential uses of an underutilized multipurpose tree in Ethiopia. *Afr. J. Plant Sci.* **2013**, *7*, 468–475. [[CrossRef](#)]
272. Rubatzky, V.E.; Yamaguchi, M. *World Vegetables: Principles, Production, and Nutritive Values*, 2nd ed.; Springer: New York, NY, USA, 1997; ISBN 978-1-4613-7756-6.
273. Bhat, R.; Karim, A. Exploring the nutritional potential of wild and underutilized legumes. *Compr. Rev. Food Sci. Food Saf.* **2009**, *8*, 305–331. [[CrossRef](#)]
274. Ajayi, I.A.; Oderinde, R.A.; Kajogbola, D.O.; Uponi, J.I. Oil content and fatty acid composition of some underutilized legumes from Nigeria. *Food Chem.* **2006**, *99*, 115–120. [[CrossRef](#)]
275. Grollier, C.; Debien, C.; Dornier, M.; Reynes, M. Principales caractéristiques et voies de valorisation du tamarin. *Fruits* **1998**, *53*, 271–280.
276. Van Der Stege, C.; Prehlsler, S.; Hartl, A.; Vogl, C.R. Tamarind (*Tamarindus indica* L.) in the traditional West African diet: Not just a famine food. *Fruits* **2011**, *66*, 171–185. [[CrossRef](#)]
277. Gulisano, A.; Alves, S.; Martins, J.N.; Trindade, L.M. Genetics and breeding of *Lupinus mutabilis*: An emerging protein crop. *Front. Plant Sci.* **2019**, *10*, 1385. [[CrossRef](#)]
278. Porch, T.G.; Cichy, K.; Wang, W.; Brick, M.; Beaver, J.S.; Santana-Morant, D.; Grusak, M.A. Nutritional composition and cooking characteristics of tepary bean (*Phaseolus acutifolius* Gray) in comparison with common bean (*Phaseolus vulgaris* L.). *Genet. Resour. Crop. Evol.* **2016**, *64*, 935–953. [[CrossRef](#)]
279. Vadivel, V.; Janardhanan, K. Nutritional and anti-nutritional composition of velvet bean: An under-utilized food legume in south India. *Int. J. Food Sci. Nutr.* **2000**, *51*, 279–287. [[CrossRef](#)] [[PubMed](#)]
280. Amoo, I.; Atasié, V.; Kolawole, O. Proximate composition, nutritionally valuable minerals, protein functional properties and anti-nutrient contents of *Mucuna preta*, *Mucuna ghana* and *Mucuna veracruz* Mottle. *Pak. J. Nutr.* **2009**, *8*, 1204–1208. [[CrossRef](#)]
281. Anugroho, F.; Kitou, M.; Kinjo, K.; Kobashigawa, N. Growth and nutrient accumulation of winged bean and velvet bean as cover crops in a subtropical region. *Plant Prod. Sci.* **2010**, *13*, 360–366. [[CrossRef](#)]
282. Arivalagan, M.; Prasad, T.; Singh, H.; Kumar, A. Variability in biochemical and mineral composition of *Mucuna pruriens* (L.) DC—An underutilized tropical legume. *Legum. Res. Int. J.* **2014**, *37*, 483. [[CrossRef](#)]
283. Balogun, I.; Olatidoye, O. Chemical composition and nutritional evaluation of velvet bean seeds (*Mucuna utilis*) for domestic consumption and industrial utilization in Nigeria. *Pak. J. Nutr.* **2012**, *11*, 116–122. [[CrossRef](#)]
284. Bhat, R.; Sridhar, K.; Young, C.-C.; Bhagwath, A.A.; Ganesh, S. Composition and functional properties of raw and electron beam-irradiated *Mucuna pruriens* seeds. *Int. J. Food Sci. Technol.* **2008**, *43*, 1338–1351. [[CrossRef](#)]
285. Chikagwa-Malunga, S.; Adesogan, A.; Sollenberger, L.; Badinga, L.; Szabo, N.; Littell, R. Nutritional characterization of *Mucuna pruriens*: 1. Effect of maturity on the nutritional quality of botanical fractions and the whole plant. *Anim. Feed. Sci. Technol.* **2009**, *148*, 34–50. [[CrossRef](#)]
286. Noman, A.; Hoque, M.; Haque, M.; Pervin, F.; Karim, M. Nutritional and anti-nutritional components in *Pachyrhizus erosus* L. tuber. *Food Chem.* **2007**, *102*, 1112–1118. [[CrossRef](#)]

287. Aremu, C.; Ige, S.A.; Ibirinde, D.; Raji, I.; Abolusoro, S.; Ajiboye, B.; Obaniyi, S.; Adekiya, A.; Asaleye, A. Assessing yield stability in African yam bean (*Sphenostylis stenocarpa*) performance using year effect. *Open Agric.* **2020**, *5*, 202–212. [[CrossRef](#)]
288. Skylas, D.J.; Paull, J.G.; Hughes, D.G.D.; Gogel, B.; Long, H.; Williams, B.; Mundree, S.; Blanchard, C.; Quail, K.J. Nutritional and anti-nutritional seed-quality traits of faba bean (*Vicia faba*) grown in south Australia. *Crop. Pasture Sci.* **2019**, *70*, 463–472. [[CrossRef](#)]
289. Wood, J.A.; Tan, H.-T.; Collins, H.M.; Yap, K.; Khor, S.F.; Lim, W.L.; Xing, X.; Bulone, V.; Burton, R.A.; Fincher, G.B.; et al. Genetic and environmental factors contribute to variation in cell wall composition in mature desi chickpea (*Cicer arietinum* L.) cotyledons. *Plant Cell Environ.* **2018**, *41*, 2195–2208. [[CrossRef](#)]
290. Wood, J.A.; Knights, E.J.; Campbell, G.M.; Choct, M. Differences between easy- and difficult-to-mill chickpea (*Cicer arietinum* L.) genotypes. Part II: Protein, lipid and mineral composition. *J. Sci. Food Agric.* **2013**, *94*, 1446–1453. [[CrossRef](#)]
291. Wood, J.A.; Knights, E.J.; Campbell, G.M.; Choct, M. Differences between easy- and difficult-to-mill chickpea (*Cicer arietinum* L.) genotypes. Part III: Free sugar and non-starch polysaccharide composition. *J. Sci. Food Agric.* **2013**, *94*, 1454–1462. [[CrossRef](#)]
292. Celmeli, T.; Sari, H.; Canci, H.; Sari, D.; Adak, A.; Eker, T.; Tokar, C. The nutritional content of common bean (*Phaseolus vulgaris* L.) landraces in comparison to modern varieties. *Agronomy* **2018**, *8*, 166. [[CrossRef](#)]
293. Paredes, C.M.; Becerra, V.V.; Tay, J.U. Inorganic nutritional composition of common bean (*Phaseolus vulgaris* L.) genotypes race Chile. *Chil. J. Agric. Res.* **2009**, *69*, 486–495. [[CrossRef](#)]
294. Batista, K.A.; Prudêncio, S.H.; Fernandes, K.F. Changes in the functional properties and antinutritional factors of extruded hard-to-cook common beans (*Phaseolus vulgaris*, L.). *J. Food Sci.* **2010**, *75*, C286–C290. [[CrossRef](#)]
295. Thavarajah, D.; Thavarajah, P.; Sarker, A.; Materne, M.; Vandemark, G.; Shrestha, R.; Idrissi, O.; Hacikamiloglu, O.; Bucak, B.; Vandenberg, A. A global survey of effects of genotype and environment on selenium concentration in lentils (*Lens culinaris* L.): Implications for nutritional fortification strategies. *Food Chem.* **2011**, *125*, 72–76. [[CrossRef](#)]
296. Thavarajah, D.; Thavarajah, P.; See, C.-T.; Vandenberg, A. Phytic acid and Fe and Zn concentration in lentil (*Lens culinaris* L.) seeds is influenced by temperature during seed filling period. *Food Chem.* **2010**, *122*, 254–259. [[CrossRef](#)]
297. Summerfield, R.J.; Muehlbauer, F.J. Mineral nutrient composition of lentil seeds 1. *Commun. Soil Sci. Plant Anal.* **1982**, *13*, 317–333. [[CrossRef](#)]
298. Sujak, A.; Kotlarz, A.; Strobel, W. Compositional and nutritional evaluation of several lupin seeds. *Food Chem.* **2006**, *98*, 711–719. [[CrossRef](#)]
299. Uhumwangho, E.S.; Omoregie, E.S. Evaluation of nutritive, antinutritive and mineral content of *Tetracarpidium conophorum* (African walnut) seed oil at different stages of fruit maturation. *Saudi J. Life Sci.* **2017**, *2*, 210–216.
300. Ayodeji, A.E.; Aliyu, N. *Tetracarpidium conophorum* (African walnut) Hutch. & Dalziel: Ethnomedicinal uses and its therapeutic activities. *J. Med. Plants Econ. Dev.* **2018**, *2*, 10. [[CrossRef](#)]
301. Nergiz, C.; Yalcin, H.; Yildiz, H. Some analytical characters of cottonseed varieties grown in Turkey. *Grasas Aceites* **1997**, *48*, 411–414. [[CrossRef](#)]
302. Renuka, C.K.; Kumarmath, P.S.; Kadakol, J.C.; Hosamani, S.V. Chemical composition and antinutritional factors in different parts and whole cotton (*Gossypium hirsutum*) plant. *Karnataka J. Agric. Sci.* **2005**, *18*, 114–117.
303. Bellaloui, N.; Turley, R.; Stetina, S. Cottonseed protein, oil, and minerals in cotton (*Gossypium hirsutum* L.) lines differing in curly leaf morphology. *Plants* **2021**, *10*, 525. [[CrossRef](#)]
304. Akobundu, E.N.T.; Cherry, J.P.; Simmons, J.G. Chemical, functional, and nutritional properties of Egusi (*Colocynthis citrullus* L.) seed protein products. *J. Food Sci.* **1982**, *47*, 829–835. [[CrossRef](#)]
305. Souci, S.W.; Fachmann, W.; Kraut, H. *Food Composition and Nutrition Tables*; MedPharm GmbH Scientific Publishing: Boca Raton, FL, USA, 2008; ISBN 978-3-8047-5038-8.
306. Bell, J.M.; Rakow, G.; Downey, R.K. Mineral composition of oil-free seeds of *Brassica napus*, *B. rapa* and *B. juncea* as affected by location and year. *Can. J. Anim. Sci.* **1999**, *79*, 405–408. [[CrossRef](#)]
307. Lajolo, F.M.; Marquez, U.M.L.; Filisetti-Cozzi, T.M.C.C.; McGregor, D.I. Chemical composition and toxic compounds in rapeseed (*Brassica napus*, L.) cultivars grown in Brazil. *J. Agric. Food Chem.* **1991**, *39*, 1933–1937. [[CrossRef](#)]
308. Reddy, N.R.; Pierson, M.D.; Sathe, S.K.; Salunkhe, D.K. Dry bean tannins: A review of nutritional implications. *J. Am. Oil Chem. Soc.* **1985**, *62*, 541–549. [[CrossRef](#)]
309. Coulibaly, M.; Kouamé, C.A.; N'Dri, D.Y.; Kouassi, N.K.; Pereko, K.K.A.; Amani, G.N. Effect of post-harvest traditional technologies on the nutrient content and antioxidant compounds of defatted flours from *Ricinodendron heudelottii* (Baill. Pierre ex Pax) seed kernels. *Technologies* **2018**, *6*, 37. [[CrossRef](#)]
310. Nyananyo, B.L.; Nyingifa, A.L. Phytochemical Investigation on the seed of *Sphenostylis stenocarpa* (Hochst Ex A. Rich.) harms (family Fabaceae). *J. Appl. Sci. Environ. Manag.* **2011**, *15*, 419–423.
311. Ellison, S.L. Carotenoids: Physiology. In *Encyclopedia of Food and Health*; Caballero, B., Finglas, P.M., Toldrá, F., Eds.; Academic Press: Oxford, UK, 2016; pp. 670–675. ISBN 978-0-12-384953-3.
312. Okafor, J.; Ani, J.C.; Okafor, G.I. Effect of processing methods on qualities of Bambara groundnut (*Voandzeia subterranea* (L.) Thouars) flour and their acceptability in extruded snacks. *Am. J. Food Technol.* **2014**, *9*, 350–359. [[CrossRef](#)]
313. Unigwe, A.; Doria, E.; Adebola, P.; Gerrano, A.S.; Pillay, M. Anti-nutrient analysis of 30 Bambara groundnut (*Vigna subterranea*) accessions in South Africa. *J. Crop Improv.* **2017**, *32*, 208–224. [[CrossRef](#)]

314. Camelo, S.; Torres, V.; Diaz, M.F. Multivariate analysis of the anti-nutritional factors of the seasonal legumes' grains. *Cuba. J. Agric. Sci.* **2008**, *42*, 339–341.
315. Franco, O.L.; dos Santos, R.C.; Batista, J.A.; Mendes, A.C.M.; de Araújo, M.A.M.; Monnerat, R.G.; Grossi-De-Sá, M.F.; de Freitas, S.M. Effects of black-eyed pea trypsin/chymotrypsin inhibitor on proteolytic activity and on development of *Anthonomus grandis*. *Phytochemistry* **2003**, *63*, 343–349. [[CrossRef](#)]
316. Grela, E.; Studziński, T.; Matras, J. Antinutritional factors in seeds of *Lathyrus sativus* cultivated in Poland. *Lathyrus Lathyrism Newsl.* **2001**, *2*, 101–104.
317. Campbell, C.G. *Grass Pea: Lathyrus sativus L.: Promoting the Conservation and Use of Underutilized and Neglected Crops*; IPGRI: Rome, Italy, 1997; ISBN 978-92-9043-341-5.
318. Girma, D.; Korbu, L. Genetic improvement of grass pea (*Lathyrus sativus*) in Ethiopia: An unfulfilled promise. *Plant Breed.* **2012**, *131*, 231–236. [[CrossRef](#)]
319. Lambein, F.; Travella, S.; Kuo, Y.-H.; Van Montagu, M.; Heijde, M. Grass pea (*Lathyrus sativus* L.): Orphan crop, nutraceutical or just plain food? *Planta* **2019**, *250*, 821–838. [[CrossRef](#)] [[PubMed](#)]
320. Abiola, C.; Oyetayo, V.O. Proximate and anti-nutrient contents of Kersting's groundnut (*Macrotyloma geocarpum*) subjected to different fermentation methods. *Br. Microbiol. Res. J.* **2015**, *10*, 1–10. [[CrossRef](#)]
321. Betancur-Ancona, D.; Gallegos-Tintoré, S.; Delgado-Herrera, A.; Pérez-Flores, V.; Ruelas, A.C.; Chel-Guerrero, L. Some physicochemical and antinutritional properties of raw flours and protein isolates from *Mucuna pruriens* (velvet bean) and *Canavalia ensiformis* (Jack bean). *Int. J. Food Sci. Technol.* **2008**, *43*, 816–823. [[CrossRef](#)]
322. Laurena, A.C.; Revilleza, M.R.; Mendoza, E.M.T. Polyphenols, phytate, cyanogenic glycosides, and trypsin inhibitor activity of several Philippine indigenous food legumes. *J. Food Compos. Anal.* **1994**, *7*, 194–202. [[CrossRef](#)]
323. Shimelis, E.A.; Rakshit, S.K. Effect of processing on antinutrients and in vitro protein digestibility of kidney bean (*Phaseolus vulgaris* L.) varieties grown in East Africa. *Food Chem.* **2007**, *103*, 161–172. [[CrossRef](#)]
324. Drew, M.D.; Borgeson, T.L.; Thiessen, D.L. A review of processing of feed ingredients to enhance diet digestibility in finfish. *Anim. Feed Sci. Technol.* **2007**, *138*, 118–136. [[CrossRef](#)]
325. Nyembwe, P.; Minnaar, A.; Duodu, K.G.; de Kock, H.L. Sensory and physicochemical analyses of roasted Marama beans [*Tylosema esculentum* (Burchell) A. Schreiber] with specific focus on compounds that may contribute to bitterness. *Food Chem.* **2015**, *178*, 45–51. [[CrossRef](#)] [[PubMed](#)]
326. Mubarak, A. Nutritional composition and antinutritional factors of mung bean seeds (*Phaseolus aureus*) as affected by some home traditional processes. *Food Chem.* **2005**, *89*, 489–495. [[CrossRef](#)]
327. Berger, J.; Robertson, L.; Cocks, P. Agricultural potential of Mediterranean grain and forage legumes: 2) Anti-nutritional factor concentrations in the genus *Vicia*. *Genet. Resour. Crop. Evol.* **2003**, *50*, 201–212. [[CrossRef](#)]
328. Lolas, G.M.; Markakis, P. The phytase of navy beans (*Phaseolus vulgaris*). *J. Food Sci.* **1977**, *42*, 1094–1097. [[CrossRef](#)]
329. Richardson, J. The Effects of Pretreatment Conditions and Micronization on the Anti-Nutritional Factors, Cookability, and Microorganisms in Navy Beans (*Phaseolus vulgaris* L.). Ph.D. Thesis, University of Manitoba, Winnipeg, MB, Canada, 2003.
330. Kelkar, S.; Siddiq, M.; Harte, J.; Dolan, K.; Nyombaire, G.; Suniaga, H. Use of low-temperature extrusion for reducing phytohemagglutinin activity (PHA) and oligosaccharides in beans (*Phaseolus vulgaris* L.) cv. Navy and Pinto. *Food Chem.* **2012**, *133*, 1636–1639. [[CrossRef](#)]
331. Saxena, K.B.; Kumar, R.V.; Sultana, R. Quality nutrition through pigeonpea—A review. *Health* **2010**, *2*, 1335–1344. [[CrossRef](#)]
332. Kaur, D.; Kapoor, A. Nutrient composition and antinutritional factors of rice bean (*Vigna umbellata*). *Food Chem.* **1992**, *43*, 119–124. [[CrossRef](#)]
333. Hory, H.-D.; Belitz, H.-D. Vergleichende Untersuchungen der tryptinspezifischen reaktiven Zentren verschiedener *Phaseolus coccineus* und *Phaseolus vulgaris*-Inhibitoren. *Eur. Food Res. Technol.* **1976**, *162*, 341–347. [[CrossRef](#)]
334. Morgan, M.; Manen, J. Lectin variability in *Phaseolus coccineus*. *Phytochemistry* **1985**, *24*, 1981–1985. [[CrossRef](#)]
335. Vadivel, V.; Janardhanan, K. The nutritional and antinutritional attributes of sword bean [*Canavalia gladiata* (Jacq.) DC.]: An under-utilized tribal pulse from south India. *Int. J. Food Sci. Technol.* **2004**, *39*, 917–926. [[CrossRef](#)]
336. De Caluw, E.; Halamov, K.; Van Damme, P. *Tamarindus indica* L.: A review of traditional uses, phytochemistry and pharmacology. *Afr. Focus* **2010**, *23*, 53–83. [[CrossRef](#)]
337. Suneja, Y.; Kaur, S.; Gupta, A.K.; Kaur, N. Levels of nutritional constituents and antinutritional factors in black gram (*Vigna mungo* L. Hepper). *Food Res. Int.* **2011**, *44*, 621–628. [[CrossRef](#)]
338. Misra, L.; Wagner, H. Alkaloidal constituents of *Mucuna pruriens* seeds. *Phytochemistry* **2004**, *65*, 2565–2567. [[CrossRef](#)] [[PubMed](#)]
339. Adepoju, G.K.A.; Odubena, O.O. Effect of *Mucuna pruriens* on some haematological and biochemical parameters. *J. Med. Plants Res.* **2009**, *3*, 073–076.
340. Gurumoorthi, P.; Janardhanan, K.; Kalavathy, G. Improving nutritional value of velvet bean, *Mucuna pruriens* (L.) DC. Var. utilis (Wall.Ex.Wight) L. H. Bailey, an under-utilized pulse, using microwave technology. *Indian J. Tradit. Knowl.* **2013**, *12*, 677–681.
341. Varga, E.; Varga, M. Development and validation of an LC-MS/MS method for the analysis of L-DOPA in oat. *Acta Biol. Szeged.* **2014**, *58*, 133–137.
342. Chandrashekharaiah, K.S. Studies on proteinase inhibitors from the seeds of *Mucuna utilis*. *Res. J. Chem. Environ.* **2018**, *22*, 23–27.
343. Kotaru, M.; Ikeuchi, T.; Yoshikawa, H.; Ibuki, F. Investigations of antinutritional factors of the winged bean (*Psophocarpus tetragonolobus*). *Food Chem.* **1987**, *24*, 279–286. [[CrossRef](#)]

344. Shet, M.S.; Madaiah, M. Distribution of lectin activity at different stages in the tissues of winged bean (*Psophocarpus tetragonolobus* (L.) DC). *Plant Sci.* **1987**, *53*, 161–165. [[CrossRef](#)]
345. Topal, N.; Bozoğlu, H. Determination of L-Dopa (L-3, 4-Dihydroxyphenylalanine) content of some Faba bean (*Vicia faba* L.) genotypes. *J. Agric. Sci.* **2016**, *22*, 145–151.
346. El-Adawy, T.A. Nutritional composition and antinutritional factors of chickpeas (*Cicer arietinum* L.) undergoing different cooking methods and germination. *Mater. Veg.* **2002**, *57*, 83–97. [[CrossRef](#)]
347. Sathe, S.K.; Deshpande, S.S.; Reddy, N.R.; Goll, D.E.; Salunkhe, D.K. Effects of germination on proteins, raffinose oligosaccharides, and antinutritional factors in the great northern beans (*Phaseolus vulgaris* L.). *J. Food Sci.* **1983**, *48*, 1796–1800. [[CrossRef](#)]
348. Savage, G.; Deo, S. The nutritional value of peas (*Pisum sariurn*): A literature review. *Nutr. Abstr. Rev. Ser. A* **1989**, *59*, 65–87.
349. Hefnawy, T. Effect of processing methods on nutritional composition and anti-nutritional factors in lentils (*Lens culinaris*). *Ann. Agric. Sci.* **2011**, *56*, 57–61. [[CrossRef](#)]
350. Thavarajah, P.; Thavarajah, D.; Vandenberg, A. Low phytic acid lentils (*Lens culinaris* L.): A potential solution for increased micronutrient bioavailability. *J. Agric. Food Chem.* **2009**, *57*, 9044–9049. [[CrossRef](#)]
351. Getachew, P.; Umata, M.; Retta, N.; Bekele, T.; Haki, G.D. Proximate composition and anti-nutritional factors of traditionally processed white lupine (*Lupinus albus* L.) Fabaceae, grown in Ethiopia. *Ethiop. J. Biol. Sci.* **2012**, *11*, 133–146.
352. Nigussie, Z. Contribution of white lupin (*Lupinus albus* L.) for food security in northwestern Ethiopia: A review. *Asian J. Plant Sci.* **2012**, *11*, 200–205. [[CrossRef](#)]
353. Janßen, G.; Ordon, F.; Jürgens, H.-U. Effects of temperature on the alkaloid content of seeds of *Lupinus angustifolius* Cultivars. *J. Agron. Crop. Sci.* **2009**, *195*, 172–177. [[CrossRef](#)]
354. Boye, J.; Ribereau, S. Assessing compositional differences in soy products and impacts on health claims. In *Soybean and Nutrition*; IntechOpen: London, UK, 2011. [[CrossRef](#)]